

The aptasensors developed showed reproducibility of the R_{et} value which ranges from 4.5% for single measurements performed with six aptasensors for 10 nM OTA to 7.5% for a series of six measurements with the same aptasensor within a week operation. Commonly, the metrological characteristics obtained with the same sensor appear better than those of different sensors. In this case, the decrease of reproducibility is related to insufficient recovery of the aptasensor response. Treating the aptasensor with 0.1 M EDTA solution and 0.1 M NaCl did not lead to full recovery on the EIS characteristics. Instead, the response tends to decrease down to 80% recovery in six consecutive measurements. Meanwhile the storage of the aptasensors prepared prior to their application in dry conditions at 4 °C for at least two weeks did not alter the signal, while its variation increased during the storage period by 1.5–2.0 fold. For these reasons, the developed aptasensors can be recommended for a single use without any regeneration after their contact with the sample. Taking into account very low amounts of modifiers as well as simple preparation protocol, this does not lead significant increase in the measurement cost. As regards the Au/Boltorn H30[®] suspension, it can be stored at 4 °C for at least one month without significant changes of the characteristics of the aptasensor prepared from it. Moderate improvement of the signal reproducibility can be achieved by sonication of the suspension prior to its use for 5–10 min. The procedure does not lead significant changes in the distribution of the Au nanoparticle size, but increases the uniformity of the sensing layer.

3.4. Selectivity and Real Sample Assay

The selectivity of the developed aptasensor was estimated under similar experimental conditions using an ochratoxin B standard solution. The slope of the calibration curve obtained in the concentration range from 1.0 nM to 100 nM was 25 k Ω /log c , or three times lower than that of OTA. The LOD of 1.0 nM makes it possible to detect at least 50-times higher concentration of the target analyte without any interference and up to 50 nM with less than 10% deviation of the result. The maximal difference in the signals toward OTA and ochratoxin B were achieved for the OTA concentration of 10 nM. This is quite acceptable for direct detection of OTA in foodstuffs.

The application of Au nanoparticles as an aptamer carrier can interfere with some biological compounds, e.g., amino acids or thiols which are frequently present in the samples tested. However, no significant influence of 0.1 mM glycine, alanine, phenylalanine and cysteine added prior to or together with 10 nM OTA on charge transfer resistance was observed. The stability of the aptasensor signal can be referred to a strong interaction of Au nanoparticles with thiolated aptamer which is placed on their surface prior to contact with the sample. Rather dense coverage of the carrier surface with aptamer molecules prevents amino acids and thiols from their reaction with golden nanoparticles.

To confirm the prospects of the aptasensor in real sample assay, it was tested on the spiked samples of light and dark beer (“White Bear” and “Žatecký Gus Černý”, respectively). Prior to OTA spiking, the beer samples were boiled for 15 min until foaming stopped and then mixed with distilled water to their initial volume. The signal was measured in the conditions described for standard solutions. The recovery of about 70% for light beer and 78% for dark beer was obtained for six measurements with 5 nM OTA. Some losses of the analyte can be related to the OTA adsorption on solid particles remained in the beer. In HPLC experiments they are removed by filtration prior to OTA addition.

The direct detection of OTA in undiluted beer accelerates testing and makes it possible to detect the OTA quantities below the maximal admissible levels established for foodstuffs [13,14].

4. Conclusions

In this work, an impedimetric aptasensor has been developed for OTA detection on the base of novel aptamer carrier based on Au nanoparticles suspended in the dendrimeric hydrophilic polymer Boltorn H30[®]. Measurements of electrochemical properties of the modifier confirmed the high activity of Au nanoparticles in the electron transduction as well as improvement of the aptasensor characteristics in comparison with Boltorn H30[®] and naked electrode. The use of the polymeric form of Neutral Red and thiolated aptamer against OTA made it possible to develop an easy protocol of aptamer immobilization and ensured the high sensitivity of the response. A LOD of 0.02 nM achieved under optimal conditions of biolayer assembly is lower than that of similar aptasensors with other signal transduction principles.

Acknowledgments

The financial support of Russian Foundation for Basic Research (grant 11-03-00381) is gratefully acknowledged. Tibor Hianik announces fellowship of the Russian President Program for young scientists (SP-1337.2012.4). Tibor Hianik is grateful to Slovak Research and Development Agency (contract No. APVV-0410-10), VEGA (project No. 1/0785/12) and to Centre of Excellence SAS for Functionalized Multiphase Materials (FUN-MAT) for financial support.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Abarca, M.; Accensi, F.; Bragulat, M.; Castella, G.; Cabañes, F. *Aspergillus carbonarius* as the main source of ochratoxin A contamination in dried vine fruits from the Spanish market. *J. Food Prot.* **2007**, *66*, 54–56.
2. Krska, R.; Schubert-Ullrich, P.; Molinelli, A.; Sulyok, M.; Macdonald, S.; Crews, C. Mycotoxin analysis: An update. *Food Addit. Contam.* **2008**, *25*, 152–163.
3. Araguás, C.; González-Peñas, E.; López de Cerain, A. Study on ochratoxin A in cereal derived products from Spain. *Food Chem.* **2005**, *92*, 459–464.
4. Duarte, S.C.; Pena, A.; Lino, C.M. A review on ochratoxin A occurrence and effects of processing of cereal and cereal derived food products. *Food Microbiol.* **2010**, *27*, 187–198.
5. Covarelli, L.; Beccari, G.; Marini, A.; Tosi, L. A review on the occurrence and control of ochratoxigenic fungal species and ochratoxin A in dehydrated grapes, non-fortified dessert wines and dried vine fruit in the Mediterranean area. *Food Control* **2012**, *26*, 347–356.
6. Blesa, J.; Soriano, J.M.; Moltó, J.C.; Mañes. Factors affecting the presence of ochratoxin A in wines. *J. Crit. Rev. Food Sci. Nutr.* **2006**, *46*, 473–478.
7. Mantle, P.G. Ochratoxin A in coffee. *J. Food Mycol.* **1998**, *1*, 63–65.

8. Kuiper-Goodman, T.; Scoot, P.M. Risk assessment of ochratoxin A: An update. *Food Addit. Contam.* **1996**, *13*, 53–57.
9. Denli, M.; Perez, J.F. Ochratoxins in feed, a risk for animal and human health: Control strategies. *Toxins* **2010**, *2*, 1065–1077.
10. Paterson, R.R.M.; Lima, N. Toxicology of mycotoxins. *Mol. Clin. Environ. Toxicol. Exp. Suppl.* **2010**, *100*, 31–63.
11. Lüthe, A.; Hildebrand, H.; Bach, U.; Dingermann, T.; Ahr, H.-J. A new approach to studying ochratoxin A (OTA)-induced nephrotoxicity: Expression profiling in vivo and in vitro employing cDNA microarrays. *Toxicol. Sci.* **2003**, *73*, 315–328.
12. El Khoury, A.; Atoui, A. Ochratoxin A: General overview and actual molecular status. *Toxins* **2010**, *2*, 461–493.
13. Commission regulation (EC) No 466/2001 of 8 March 2001 setting maximum levels for certain contaminants in foodstuffs. *Off. J. Eur. Commun.* **2001**, *77*, 1–13.
14. Commission regulation (EC) No 1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in foodstuffs. *Off. J. Eur. Commun.* **2006**, *364*, 5–24.
15. Kong, W.-J.; Liu, S.-Y.; Qiu, F.; Xiao, X.-H.; Yang, M.-H. Simultaneous multi-mycotoxin determination in nutmeg by ultrasound-assisted solid–liquid extraction and immunoaffinity column clean-up coupled with liquid chromatography and on-line post-column photochemical derivatization-fluorescence detection. *Analyst* **2013**, *138*, 2729–2739.
16. Mao, J.; Lei, S.; Yang, X.; Xiao, D. Quantification of ochratoxin A in red wines by conventional HPLC–FLD using a column packed with core–shell particles. *Food Control* **2013**, *32*, 505–511.
17. Wei, R.; Qiu, F.; Kong, W.; Wei, J.; Yang, M.; Luo, Z.; Qin, J.; Ma, X. Co-occurrence of aflatoxin B1, B2, G1, G2 and ochratoxin A in *Glycyrrhiza uralensis* analyzed by HPLC-MS/MS. *Food Control* **2013**, *32*, 216–221.
18. Goryacheva, I.Y.; de Saeger, S.; Lobeau, M.; Eremin, S.A.; Barna-Vetró, I.; van Peteghem, C. Approach for ochratoxin A fast screening in spices using clean-up tandem immunoassay columns with confirmation by high performance liquid chromatography-tandem mass spectrometry (HPLC–MS/MS). *Anal. Chim. Acta* **2006**, *577*, 38–45.
19. Arroyo-Manzanares, N.; Gániz-Gracia L.; García-Campaña, A.M. Determination of ochratoxin A in wines by capillary liquid chromatography with laser induced fluorescence detection using dispersive liquid-liquid microextraction. *Food Chem.* **2012**, *135*, 368–372.
20. Urusova, A.E.; Kostenko, S.N.; Sveshnikov, P.G.; Zherdev, A.V.; Dzantiev, B.B. Ochratoxin A immunoassay with surface plasmon resonance registration: Lowering limit of detection by the use of colloidal gold immunoconjugates. *Sens. Actuators B* **2011**, *156*, 343–349.
21. Yuan, J.; Deng, D.; Lauren, D.R.; Aguilar, M.-I.; Wu, Y. Surface plasmon resonance biosensor for the detection of ochratoxin A in cereals and beverages. *Anal. Chim. Acta* **2009**, *656*, 63–71.
22. Liu, X.; Yang, Z.; Zhang, Y.; Yu, R. A novel electrochemical immunosensor for ochratoxin A with hapten immobilization on thionine/gold nanoparticle modified glassy carbon electrode. *Anal. Method.* **2013**, *5*, 1481–1486.
23. Bonel, L.; Vidal, J.C.; Duato, P.; Castillo, J.R. Ochratoxin A nanostructured electrochemical immunosensors based on polyclonal antibodies and gold nanoparticles coupled to the antigen. *Anal. Method.* **2010**, *2*, 335–341.

24. Liu, X.-P.; Deng, Y.-J.; Jin, X.-Y.; Chen, L.-G.; Jiang, J.-H.; Shen, G.-L.; Yu, R.-Q. Ultrasensitive electrochemical immunosensor for ochratoxin A using gold colloid-mediated hapten immobilization. *Anal. Biochem.* **2009**, *389*, 63–68.
25. Radi, A.-E.; Muñoz-Berbel, X.; Lates, V.; Marty, J.-L. Label-free impedimetric immunosensor for sensitive detection of ochratoxin A. *Biosens. Bioelectron.* **2009**, *24*, 1888–1892.
26. Prieto-Simón, B.; Karube, I.; Saiki, H. Sensitive detection of ochratoxin A in wine and cereals using fluorescence-based immunosensing. *Food Chem.* **2012**, *135*, 1323–1329.
27. Li, T.; Job, E.-J.; Kim, M.-G. A label-free fluorescence immunoassay system for the sensitive detection of the mycotoxin, ochratoxin A. *Chem. Commun.* **2012**, *48*, 2304–2306.
28. Huang, B.; Xiao, H.; Zhang, J.; Zhang, L.; Yang, H.; Zhang, Y.; Jin, J. Dual-label time-resolved fluoroimmunoassay for simultaneous detection of aflatoxin B1 and ochratoxin A. *Arch. Toxicol.* **2009**, *83*, 619–624.
29. Meulenberg, E.P. Immunochemical methods for ochratoxin A detection: A review. *Toxins* **2012**, *4*, 244–266.
30. Oliveira, S.C.B.; Diculescu, V.C.; Palleschi, G.; Compagnone, D.; Oliveira-Brett, A.M. Electrochemical oxidation of ochratoxin A at a glassy carbon electrode and in situ evaluation of the interaction with deoxyribonucleic acid using an electrochemical deoxyribonucleic acid-biosensor. *Anal. Chim. Acta* **2007**, *588*, 283–291.
31. Mukhopadhyay, R. Aptamers are ready for the spotlight. *Anal. Chem.* **2005**, *77*, 115A–118A.
32. Tuerk, C.; Gold, L. Systematic evolution of ligands by exponential enrichment: RNA ligands to bacteriophage T4 DNA polymerase. *Science* **1990**, *249*, 505–510.
33. Ellington, A.D.; Szostak, J.W. *In vitro* selection of RNA molecules that bind specific ligands. *Nature* **1990**, *346*, 818–822.
34. *Aptamers in Analysis*; Mascini, M., Ed.; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2009.
35. Song, K.-M.; Lee, S.; Ban, C. Aptamers and their biological applications. *Sensors* **2012**, *12*, 612–631.
36. Wang, Z.; Ma, L. Gold nanoparticle probes. *Coord. Chem. Rev.* **2009**, *253*, 1607–1618.
37. Pingarrón, J.M.; Yáñez-Sedeño, P.; González-Cortés, A. Gold nanoparticle-based electrochemical biosensors. *Electrochim. Acta* **2008**, *53*, 5848–5866.
38. Wu, J.; Chu, H.; Mei, Z.; Deng, Y.; Xue, F.; Zheng, L.; Chen, W. Ultrasensitive one-step rapid detection of ochratoxin A by the folding-based electrochemical aptasensor. *Anal. Chim. Acta* **2012**, *753*, 27–31.
39. Yang, C.; Wang, Y.; Marty, J.-L.; Yang, X. Aptamer-based colorimetric biosensing of Ochratoxin A using unmodified gold nanoparticles indicator. *Biosens. Bioelectron.* **2011**, *26*, 2724–2727.
40. Wang, Z.; Duan, N.; Hun, X.; Wu, S. Electrochemiluminescent aptamer biosensor for the determination of ochratoxin A at a gold-nanoparticles-modified gold electrode using N-(aminobutyl)-N-ethylisoluminol as a luminescent label. *Anal. Bioanal. Chem.* **2010**, *398*, 2125–2132.
41. Yin, X.-B. Functional nucleic acids for electrochemical and electrochemiluminescent sensing applications. *Trends Anal. Chem.* **2012**, *33*, 81–94.

42. Aymonier, C.; Schlotterbeck, U.; Antonietti, L.; Zacharias, P.; Thomann, R.; Tiller, J.C.; Mecking, S. Hybrids of silver nanoparticles with amphiphilic hyperbranched macromolecules exhibiting antimicrobial properties. *Chem. Commun.* **2002**, doi:10.1039/B208575E.
43. Krämer, M.; Pérignon, N.; Haag, R.; Marty, J.-D.; Thomann, R.; Lauth-de Viguier, N.; Mingotaud, C. Water-soluble dendritic architectures with carbohydrate shells for the templation and stabilization of catalytically active metal nanoparticles. *Macromolecules* **2005**, *38*, 8308–8315.
44. Yamahira, A.; Torigoe, K. Role of poly(amidoamine) dendrimers for preparing nanoparticles of gold, platinum, and silver. *Langmuir* **2000**, *16*, 2604–2608.
45. Pérignon, N.; Mingotaud, A.F.; Marty, J.D.; Lattes, I.R.; Mingotaud, C. Formation and stabilization in water of metal nanoparticles by a hyperbranched polymer chemically analogous to PAMAM dendrimers. *Chem. Mater.* **2004**, *16*, 4856–4858.
46. Wei, X.; Zhu, B.; Xu, Y. Preparation and stability of copper particles formed from the template of hyperbranched poly(amine-ester). *Colloid Polym. Sci.* **2005**, *284*, 102–107.
47. Tabuani, D.; Monticelli, O.; Chincarini, A.; Bianchini, C.; Vizza, F.; Moneti, S.; Russo, S. Palladium nanoparticles supported on hyperbranched aramids: Synthesis, characterization, and some applications in the hydrogenation of unsaturated substrates. *Macromolecule* **2003**, *36*, 4294–4301.
48. Tabuani, D.; Monticelli, O.; Komber, H.; Russo, S. Preparation and characterisation of Pd nanoclusters in hyperbranched aramid templates to be used in homogeneous catalysis. *Macromol. Chem. Phys.* **2003**, *204*, 1576–1583.
49. Ichikawa, H.; Yasui, K.; Ozawa, M.; Fujita, K. Electrical bistability of composite film comprising hyper-branched polymer and gold nanoparticle. *Synth. Met.* **2009**, *159*, 973–976.
50. Zhao, Y.; Zou, J.; Shi, W. *In situ* synthesis and characterization of lead sulfide nanocrystallites in the modified hyperbranched polyester by gamma-ray irradiation. *Mater. Sci. Eng. B* **2005**, *121*, 20–24.
51. Raveendran, P.; Goyal, A.; Blatchford, M.; Wallen, S. Stabilization and growth of silver nanocrystals in dendritic polyol dispersions. *Mater. Lett.* **2006**, *60*, 897–900.
52. Aryal, S.; Prabakaran, M.; Pilla, S.; Gong, S. Biodegradable and biocompatible multi-arm star amphiphilic block copolymer as a carrier for hydrophobic drug delivery. *Int. J. Biol. Macromol.* **2009**, *44*, 346–352.
53. Prabakaran, M.; Grailer, J.; Steeber, D.; Gong, S. Folate-conjugated amphiphilic hyperbranched block copolymers based on Boltorn[®] H40, poly(L-lactide) and poly(ethylene glycol) for tumor-targeted drug delivery. *Biomaterials* **2009**, *30*, 3009–3019.
54. Prabhakar, P.; Matharu, Z.; Malhotra, B.D. Polyaniline Langmuir–Blodgett film based aptasensor for ochratoxin A detection. *Biosens. Bioelectron.* **2011**, *26*, 4006–4011.
55. Xia, F.; Zuo, X.; Yang, R.; Xiao, X.; Kang, D.; Vallée-Bélisle, A.; Gong, G.; Yuen J.D.; Hsu, B.B.Y.; Heeger, A.J.; *et al.* Colorimetric detection of DNA, small molecules, proteins, and ions using unmodified gold nanoparticles and conjugated polyelectrolytes. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 10837–10841.
56. Evtugyn, G.; Kostyleva, V.; Sitdikov, R.; Porfireva, A.; Savelieva, M.; Stoikov, I.; Antipin, I.; Hianik T. Electrochemical aptasensor based on a macrocyclic ligand bearing Neutral Red. *Electroanalysis* **2012**, *24*, 91–100.

57. Pauliukaite, R.; Brett, C.M.A. Poly(neutral red): Electrosynthesis, characterization, and application as a redox mediator. *Electroanalysis* **2008**, *20*, 1275–1285.
58. Bard, J.; Faulkner, L.R. *Electrochemical Methods: Fundamentals and Applications*, 2nd ed.; John Wiley and Sons, Inc.: New York, NY, USA, 2001.
59. Adams, R.N. *Electrochemistry at Solid Electrodes*; Marcel-Dekker: New York, NY, USA, 1969.
60. Kuang, H.; Chen, W.; Xu, D.; Xu, L.; Zhu, Y.; Liu, L.; Chu, H.; Peng, C.; Xu, C.; Zhu, S. Fabricated aptamer-based electrochemical “signal-off” sensor of ochratoxin A. *Biosens. Bioelectron.* **2010**, *26*, 710–716.
61. Zhang, J.; Chen, J.; Zhang, X.; Zeng, Z.; Chen, M.; Wang, S. An electrochemical biosensor based on hairpin-DNA aptamer probe and restriction endonuclease for ochratoxin A detection. *Electrochem. Commun.* **2012**, *25*, 5–7.
62. Bonel, L.; Vidal, J.C.; Duato, P.; Castillo, J.R. An electrochemical competitive biosensor for ochratoxin A based on a DNA biotinylated aptamer. *Biosens. Bioelectron.* **2011**, *26*, 3254–3259.
63. Barthelmebs, L.; Hayat, A.; Limiadi, A.W.; Marty, J.-L.; Noguer, T. Electrochemical DNA aptamer-based biosensor for OTA detection, using superparamagnetic nanoparticles. *Sens. Actuators B* **2011**, *156*, 932–937.
64. Castillo, G.; Lamberti, I.; Mosiello, L.; Hianik, T. Impedimetric DNA aptasensor for sensitive detection of ochratoxin A in food. *Electroanalysis* **2012**, *24*, 512–520.
65. Evtugyn, G.; Porfireva, A.; Sitdikov, R.; Evtugyn, V.; Stoikov, I.; Antipin, I.; Hianik, T. Electrochemical aptasensor for the determination of ochratoxin A at the Au electrode modified with Ag nanoparticles decorated with macrocyclic ligand. *Electroanalysis* **2013**, *25*, 1847–1854.
66. Yin, X.B.; Xin, Y.Y.; Zhao, Y. Label-free electrochemiluminescent aptasensor with attomolar mass detection limits based on a Ru(phen)₃²⁺-double-strand DNA composite film electrode. *Anal. Chem.* **2009**, *81*, 9299–9305.

© 2013 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).