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EXPERIMENTAL PAPERS

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# Stimulation of Alpha-2 Adrenoreceptors against the Background of Hyperpolarization-Activated Current Blockade at Different Formation Stages of Cardiac Adrenergic Innervation

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**Abstract**—In different periods of postnatal development, the components of the autonomic nervous system, including specific receptors and neurotransmitters involved in the control of cardiovascular functions, are characterized by significant dynamics and variability. Although in humans and animals,  $\alpha_2$ -adrenoceptors ( $\alpha_2$ -ARs) and the hyperpolarization-activated current ( $I_f$ ) are widespread and involved in the regulation of multiple functions, their modulatory role at different stages of the formation and development of cardiac adrenergic regulation has been insufficiently studied. The present study was carried out on isolated hearts of 1-week-old, 3-week-old, and adult white rats. Heart rate (HR) and coronary flow (CF) in the Langendorff rat heart were recorded upon  $\alpha_2$ -AR activation with clonidine hydrochloride ( $10^{-6}$  M) against the background of  $I_f$  blocker ZD7288 application at  $10^{-9}$  M and  $10^{-5}$  M. In 1-week-old rat pups,  $\alpha_2$ -ARs activation evoked bidirectional changes in CF depending on  $I_f$  blocker concentration, causing its significant increase at  $10^{-9}$  M and bidirectional dynamics at  $10^{-5}$  M. In 3-week-old rat pups,  $\alpha_2$ -AR stimulation reduced CF, while against the background of preliminary  $I_f$  blockade, the CF dynamics changed to positive. In adult rats,  $\alpha_2$ -AR activation against the background of  $I_f$  blockade caused a decrease in CF. The  $\alpha_2$ -AR agonist evoked bidirectional changes in HR in 1-week-old and adult rats, causing only a significant reduction in HR in 3-week-old rat pups. Stimulation of  $\alpha_2$ -ARs against the background of  $I_f$  blockade did not alter the HR dynamics in 3-week-old rats and abolished bidirectional HR changes in adult and 1-week-old animals, causing only a decrease in HR in adults and an increase in HR in newborns. The obtained results demonstrate that  $\alpha_2$ -adrenergic cardiac regulation is modulated by  $I_f$ , with the degree and direction of the effects depending on the maturity level of cardiac sympathetic innervation.

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**Keywords:** alpha-2 adrenergic receptor, hyperpolarization-activated current, isolated heart, ontogenesis, rat

## INTRODUCTION

Heart development is a multistage process controlled by neural and humoral regulatory mechanisms. During different periods of postnatal development, the components of the autonomic nervous system, including specific receptors and neurotransmitters involved in the control of cardiovascular functions, are characterized by significant dynamics and variability [1–2]. A review of modern literature presents a wide range of results on the early formation and further development of sympathetic and parasympathetic nerve fibers in the heart [3–4]. Fluorescence microscopy does not detect sympathetic fibers in the heart tissues of rat pups during the first 21 postnatal days. The ability to uptake norepinephrine defines the functional maturity of sympathetic nerves, which is first observed on day 21 after birth, reaching the levels of mature animals by day 30 [5]. Experiments with radioactively labeled norepinephrine revealed the formation of the sympathetic nervous system on day 31 of life [5–6].

Adrenergic regulation of cardiac activity is implemented through multiple subtypes of adrenoceptors [7]. In humans and animals,  $\alpha_2$ -adrenoceptors ( $\alpha_2$ -ARs) are widely distributed in the central and peripheral nervous systems, autonomic ganglia, on the presynaptic membrane of adrenergic fibers and the postsynaptic membrane of cardiomyocytes, in vascular smooth muscles, as well as in the intestinal and renal epithelia [8–9].  $\alpha_2$ -ARs localized on presynaptic membranes are involved in modulating norepinephrine release [10], inhibiting acetylcholine release from cholinergic synapses, stimulating platelet aggregation, and vasoconstriction [11]. Activation of postsynaptic  $\alpha_2$ -ARs in the central nervous system causes a sedative effect, inhibits sympathetic activity, and reduces blood pressure [12].

Previous studies have revealed the involvement of  $\alpha_2$ -ARs in the regulation of cardiac performance parameters at different levels of organization of the organism. In *in vivo* studies,  $\alpha_2$ -AR activation causes bradycardia and reduces systolic pressure in rats; in series of *in vitro* experiments, the contractility of rat ventricular myocardial strips changes bidirectionally relative to the atria [13]. In the *ex vivo* Langendorff (i.e. isolated perfused) heart model,  $\alpha_2$ -AR activation reduces left ventricular (LV) pressure and causes

bidirectional changes in heart rate (HR) and coronary flow (CF) [14].

Alpha-2 adrenoceptors play a special role in the development and clinical course of cardiovascular diseases. Rats with a model of spontaneous arterial hypertension, demonstrate an increase in the expression of  $\alpha_2$ -AR subtypes and a dysfunction thereof, as well as a reduced efficacy of associated signaling cascades [15]. The use of an  $\alpha_2$ -AR agonist (dexmedetomidine) in hypoxia/oxygenation-induced LV dysfunction exerts a cardioprotective effect, as manifested in sympatholysis, stabilization of hemodynamics, and activation of endothelial nitric oxide synthase (eNOS) [16].

Cyclic nucleotide-dependent, hyperpolarization-activated (HCN) channels, nonselectively conducting  $\text{Na}^+$  and  $\text{K}^+$  ions, are blocked by millimolar concentrations of extracellular  $\text{Cs}^+$ , and are modulated by cyclic nucleotides, mainly cAMP [17]. The operation of HCN channels is based on the hyperpolarization-activated inward current, which is called the “funny current” ( $I_f$ ) in the cells of the sinoatrial node of the heart and  $I_h$  in the nervous system and nerve cells [17].

Since  $I_f$  discovery and description in the 1980s by DiFrancesco et al. [17], a large number of studies have been conducted to identify the involvement of these currents in the regulation of the cardiovascular system functions in the developing organism under normal and pathological conditions [18–20]. At the early stages of embryonic development, HCN channels are abundantly expressed in all parts of the heart, while by the time of birth, their expression in working cardiomyocytes decreases and remains at a low level in adults, which is essential for preventing pathological remodeling [21]. An increased expression of HCN channels has been revealed in heart failure and after myocardial infarction [22–23]. It has been established that  $I_f$  blockade causes bradycardia and reduces CF in laboratory rats of different ages [24–26]. Inhibition of  $I_f$  affects the contractile activity of the atrial and ventricular myocardium in rats at various ontogenetic periods [27].

It has been found that  $I_f$  activity is controlled by cAMP levels and is regulated by sympathetic and parasympathetic influences of the autonomic nervous system [28]. The formation of nerve fibers plays a key role in regulating ion currents of myocardial cells and the development of cardiovascular reflexes

[29].  $I_f$  and  $\alpha_2$ -ARs are present in cardiomyocytes at all stages of the formation and further development of the cardiovascular system, while  $I_f$  may be an important effector of adrenergic regulation in the developing heart.

The aim of this study was to investigate the effect of  $\alpha_2$ -AR stimulation against the background of hyperpolarization-activated current ( $I_f$ ) blockade on the parameters of an isolated rat heart at different developmental stages of cardiac sympathetic innervation.

## MATERIALS AND METHODS

*Drugs, solutions, chemical agents.* The experiments were carried out on isolated heart preparations from white outbred rats of two age groups, 1-week-old ( $n = 22$ ) and 3-week-old ( $n = 24$ ). In 1-week-old rat pups, cardiac sympathetic innervation is yet absent, while at the age of 3 weeks, its formation gets started [30]. Adult animals (20-week-old,  $n = 24$ ) with a fully formed cardiovascular regulatory system were chosen as controls.

In a study on the Langendorff isolated perfused rat heart, a Krebs-Henseleit solution for warm-blooded animals of the following composition (mmol/L) was used: NaCl 118.0, KCl 4.7, NaHCO<sub>3</sub> 25.0, MgSO<sub>4</sub> 1.2, CaCl<sub>2</sub> 2.5, KH<sub>2</sub>PO<sub>4</sub> 1.2, glucose 5.5. The solution was saturated with carbogen (95% O<sub>2</sub>, 5% CO<sub>2</sub>); the pH of the solution was maintained at 7.3–7.4 at a temperature of 37°C.

To stimulate  $\alpha_2$ -ARs, its agonist clonidine hydrochloride (Sigma-Aldrich, USA) was used at a concentration of  $10^{-6}$  M; for  $I_f$  blockade, its blocker ZD7288 (Tocris, UK) was applied at concentrations of  $10^{-9}$  M and  $10^{-5}$  M. After stabilization of the isolated heart parameters, we recorded control 1, after which added ZD7288 to the perfusate, followed (after 5 min) by clonidine hydrochloride, and recorded control 2 (ZD7288). Perfusion of the isolated heart was recorded for 20 min. Different animals were used in each series of experiments. In the graphs, the values of the isolated heart parameters, recorded at the 5th min after ZD7288 addition to the perfusate, were taken as 100% (Table 1, control 2).

*Preparing the Langendorff isolated perfused rat heart.* Rats of both sexes were anesthetized with a 25% urethane solution (800 mg/kg body weight), and the chest was opened. The heart was isolated and

flushed with an ice-cold ( $\sim 2^\circ\text{C}$ ) working Krebs–Henseleit solution. The isolated heart was fixed by the aorta to a cannula, and oxygenated working solution was supplied in the Langendorff apparatus (ADInstruments, Australia) at a temperature of 37°C. Gravitational retrograde perfusion was performed through the aorta under a constant hydrostatic pressure of 70–75 mm Hg. To monitor HR throughout the experiment, ECG was recorded using atraumatic electrodes placed directly on the heart. Changes in CF in response to pharmacological exposures were also recorded. Signals were recorded using a PowerLab 8/35 data acquisition device (ADInstruments, Australia) and LabChart-Pro software.

*Statistics.* Statistical data processing was performed in MS Excel (Microsoft Corp.) and using one-way ANOVA with the  $t$ -test for independent variables and the  $t$ -test for dependent samples in Statistica 13.0 (StatSoft, Inc.). Data are presented as  $M \pm SEM$ . Differences were considered statistically significant at  $p < 0.05$ .

## RESULTS

In 1-week-old rat pups ( $n = 8$ ), the  $\alpha_2$ -AR agonist clonidine ( $10^{-6}$  M) exerted a bidirectional effect on the HR of the isolated heart, decreasing it in some rats by 11% ( $n = 4$ ,  $p < 0.05$ ), while increasing it in the others by 30% ( $n = 4$ ,  $p < 0.05$ , Fig. 1a). In 3-week-old rat pups, clonidine ( $10^{-6}$  M) decreased HR by 31% vs. baseline values ( $n = 10$ ,  $p < 0.01$ ). In control (adult) rats,  $\alpha_2$ -AR stimulation with clonidine ( $10^{-6}$  M) caused bidirectional changes in HR: a 10% increase ( $n = 4$ ,  $p < 0.05$ ) and a 25% decrease ( $n = 4$ ,  $p < 0.05$ ) (Fig. 1a).

The  $\alpha_2$ -AR agonist clonidine ( $10^{-6}$  M) added against the background of preliminary  $I_f$  blockade with ZD7288 ( $10^{-9}$  M) caused the following events. In 1-week-old rat pups, it increased HR from  $192.4 \pm 19.3$  to  $220.8 \pm 22.8$  bpm by the 5th min of the experiment. By the 15th min, HR increased to  $225.8 \pm 21.4$  bpm ( $p < 0.05$ ), and by the 20th min, it reached  $232.9 \pm 29.9$  bpm ( $n = 7$ ,  $p < 0.05$ ) (Figs. 1b, 2b; Table 1). The overall increase in HR amounted to 20% vs. baseline values. In 3-week-old rat pups, HR decreased from  $176.7 \pm 17.6$  to  $152.9 \pm 17.0$  bpm ( $p < 0.05$ ) by the 5th min of the experiment. By the 15th min, HR decreased to  $145.0 \pm$

**Table 1.** Effect of  $\alpha_2$ -adrenoreceptor stimulation against the background of  $I_f$  blockade on heart rate and coronary flow in the Langendorff heart in rats of different ages

HEART RATE						
ZD7288 ( $10^{-9}$ M) + clonidine ( $10^{-6}$ )				ZD7288 ( $10^{-5}$ M) + clonidine ( $10^{-6}$ )		
Age	1 week ( $n = 7$ )	3 weeks ( $n = 7$ )	adult ( $n = 7$ )	1 week ( $n = 7$ )	3 weeks ( $n = 6$ )	
Control 1	216.4 $\pm$ 19.2	222.2 $\pm$ 24.2	181.2 $\pm$ 22.0	199.1 $\pm$ 9.5	246.6 $\pm$ 10.2	
1 min	204.5 $\pm$ 15.2	199.7 $\pm$ 17.3	164.9 $\pm$ 18.7	201.4 $\pm$ 13.7	246.3 $\pm$ 10.2	
5 min, control 2	192.4 $\pm$ 19.3	176.7 $\pm$ 17.6	148.0 $\pm$ 18.4	204.3 $\pm$ 14.4	218.1 $\pm$ 8.7	
5 min	220.8 $\pm$ 22.8	152.9 $\pm$ 17.0*	124.3 $\pm$ 21.2	182.2 $\pm$ 10.5	189.2 $\pm$ 10.3*	
15 min	225.8 $\pm$ 21.4*	145.0 $\pm$ 18.3**	71.2 $\pm$ 16.0*	170.9 $\pm$ 11.8*	187.7 $\pm$ 13.2*	
20 min	232.9 $\pm$ 29.9	142.6 $\pm$ 20.9**	70.9 $\pm$ 15.9*	170.3 $\pm$ 12.2*	184.2 $\pm$ 7.1**	

CORONARY FLOW						
ZD7288 ( $10^{-9}$ M) + clonidine ( $10^{-6}$ )				ZD7288 ( $10^{-5}$ M) + clonidine ( $10^{-6}$ )		
Age	1 week ( $n = 7$ )	3 weeks ( $n = 7$ )	adult ( $n = 7$ )	1 week ( $n = 7$ )		3 weeks ( $n = 6$ )
				( $n = 4$ )	( $n = 3$ )	
Control 1	0.6 $\pm$ 0.1	1.6 $\pm$ 0.1	11.6 $\pm$ 2.2	0.9 $\pm$ 0.1	1.8 $\pm$ 0.1	1.8 $\pm$ 0.1
1 min	0.6 $\pm$ 0.1	1.5 $\pm$ 0.1	11.2 $\pm$ 2.2	0.9 $\pm$ 0.1	1.7 $\pm$ 0.1	1.7 $\pm$ 0.1
5 min, control 2	0.6 $\pm$ 0.1	1.3 $\pm$ 0.1	9.3 $\pm$ 2.1	0.8 $\pm$ 0.1	1.5 $\pm$ 0.1	1.5 $\pm$ 0.1
5 min	0.6 $\pm$ 0.1	1.3 $\pm$ 0.1	7.5 $\pm$ 1.8*	0.8 $\pm$ 0.1	1.4 $\pm$ 0.1	1.5 $\pm$ 0.1
15 min	0.7 $\pm$ 0.1	1.4 $\pm$ 0.1*	6.1 $\pm$ 1.8*	1.0 $\pm$ 0.1	1.4 $\pm$ 0.1	1.9 $\pm$ 0.1
20 min	0.8 $\pm$ 0.1*	1.5 $\pm$ 0.1**	5.8 $\pm$ 1.8**	1.0 $\pm$ 0.1*	1.3 $\pm$ 0.1*	2.1 $\pm$ 0.1***

\*  $p < 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ —vs. baseline values (control 2);  $M \pm SEM$ . Control 1—control heart rate (HR) and coronary flow (CF) values before ZD7288 application. Control 2—control HR) and CF values before clonidine administration against the background of ZD7288 application.

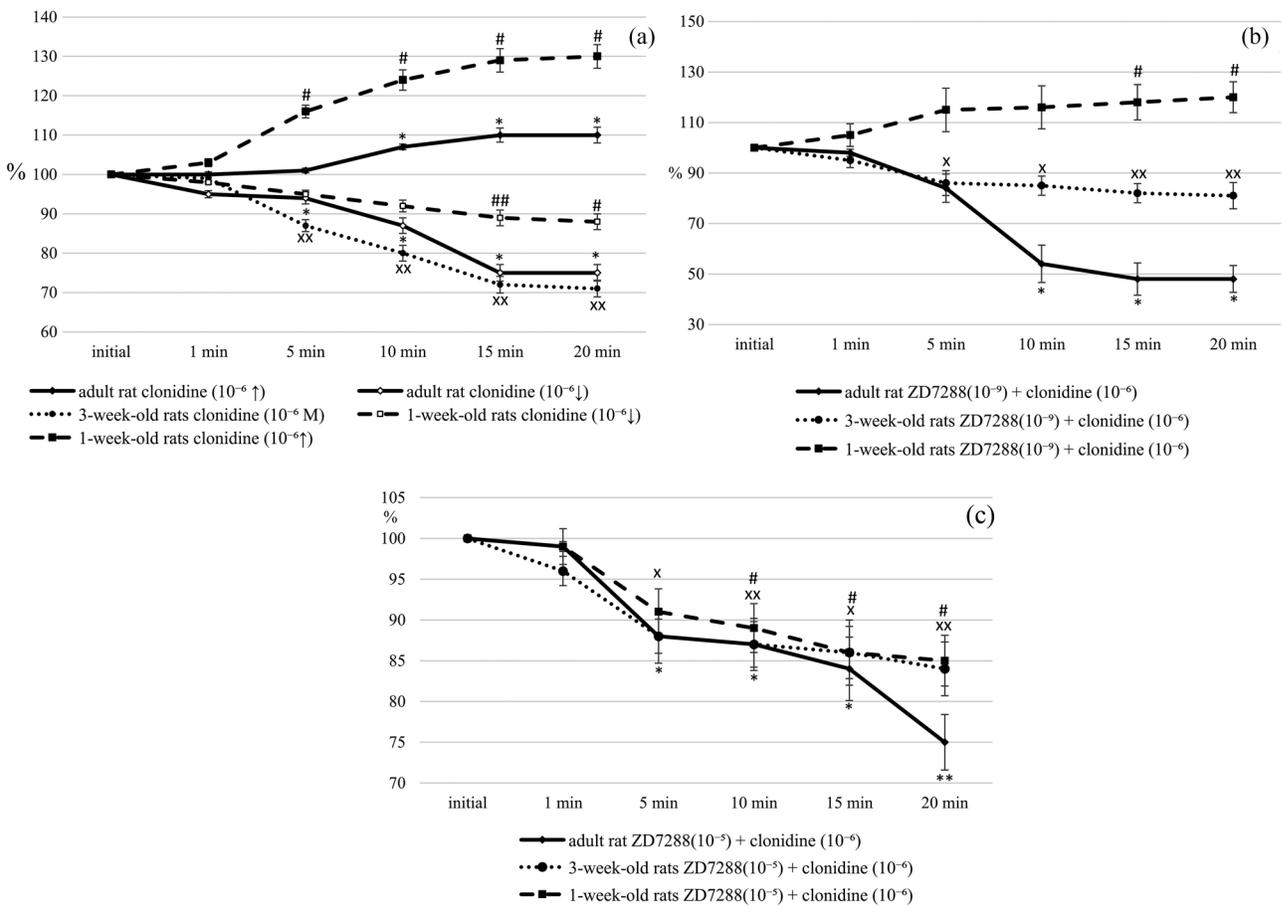
18.3 bpm ( $p < 0.01$ ), and by the 20th min, it was 142.6  $\pm$  20.9 bpm ( $n = 7$ ,  $p < 0.01$ ). The overall decrease in HR amounted to 19% (Figs. 1b, 2a). In adult rats, HR decreased by 52% ( $n = 8$ ,  $p < 0.01$ ).

The  $\alpha_2$ -AR agonist clonidine ( $10^{-6}$  M) added against the background of preliminary  $I_f$  blockade with ZD7288 at a higher concentration ( $10^{-5}$  M) exerted the following effects. In 1-week-old rat pups, HR decreased by the 5th min of the experiment from 204.3  $\pm$  14.4 to 182.2  $\pm$  10.5 bpm. By the 15th min, HR decreased to 170.9  $\pm$  11.8 bpm ( $p < 0.05$ ). By the final (20th) minute of observation, HR was 170.3  $\pm$  12.2 bpm ( $n = 7$ ,  $p < 0.05$ ) (Fig. 1c), i.e., there was an overall 15% decrease vs. baseline values. In 3-week-old rat pups, HR decreased by the 5th min of the experiment from 218.1  $\pm$  8.7 to 189.2  $\pm$  10.3 bpm

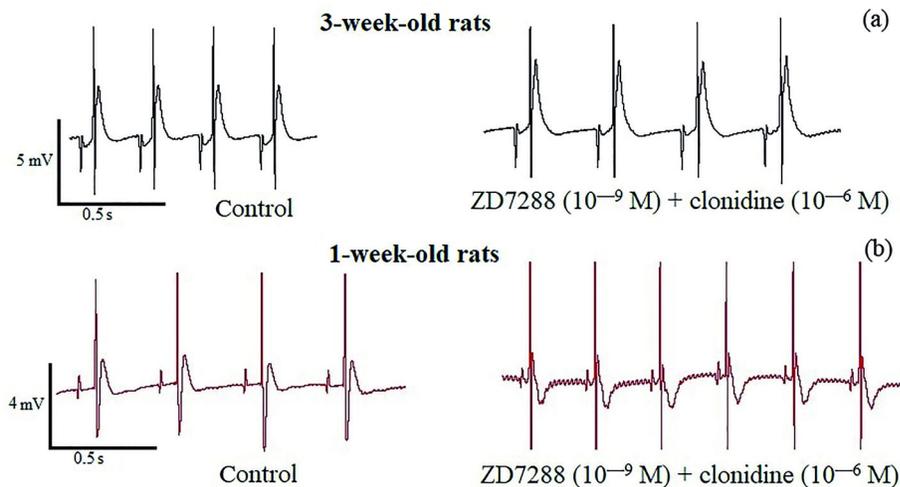
( $p < 0.05$ ). At the 15th min of observation, HR decreased to 187.7  $\pm$  13.2 bpm ( $p < 0.05$ ). By the 20th min, HR decreased to 184.2  $\pm$  7.1 bpm ( $n = 6$ ,  $p < 0.01$ ), the decrease amounted to 16% vs. baseline values. In adult rats, HR decreased by 25% ( $n = 8$ ,  $p < 0.01$ ) (Fig. 1c).

Coronary flow (CF) in 1-week-old rat pups ( $n = 8$ ) changed bidirectionally upon  $\alpha_2$ -AR activation. In some rats ( $n = 5$ ), a slight (5%) decrease in CF was observed, while in another subset of rats ( $n = 3$ ), CF increased by 4% vs. baseline values (Fig. 3a). In 3-week-old rat pups, CF decreased by 11% ( $n = 10$ ,  $p < 0.05$ ) vs. baseline values. In control (adult) rats, CF decreased by 45% ( $n = 8$ ,  $p < 0.001$ ) (Fig. 3a).

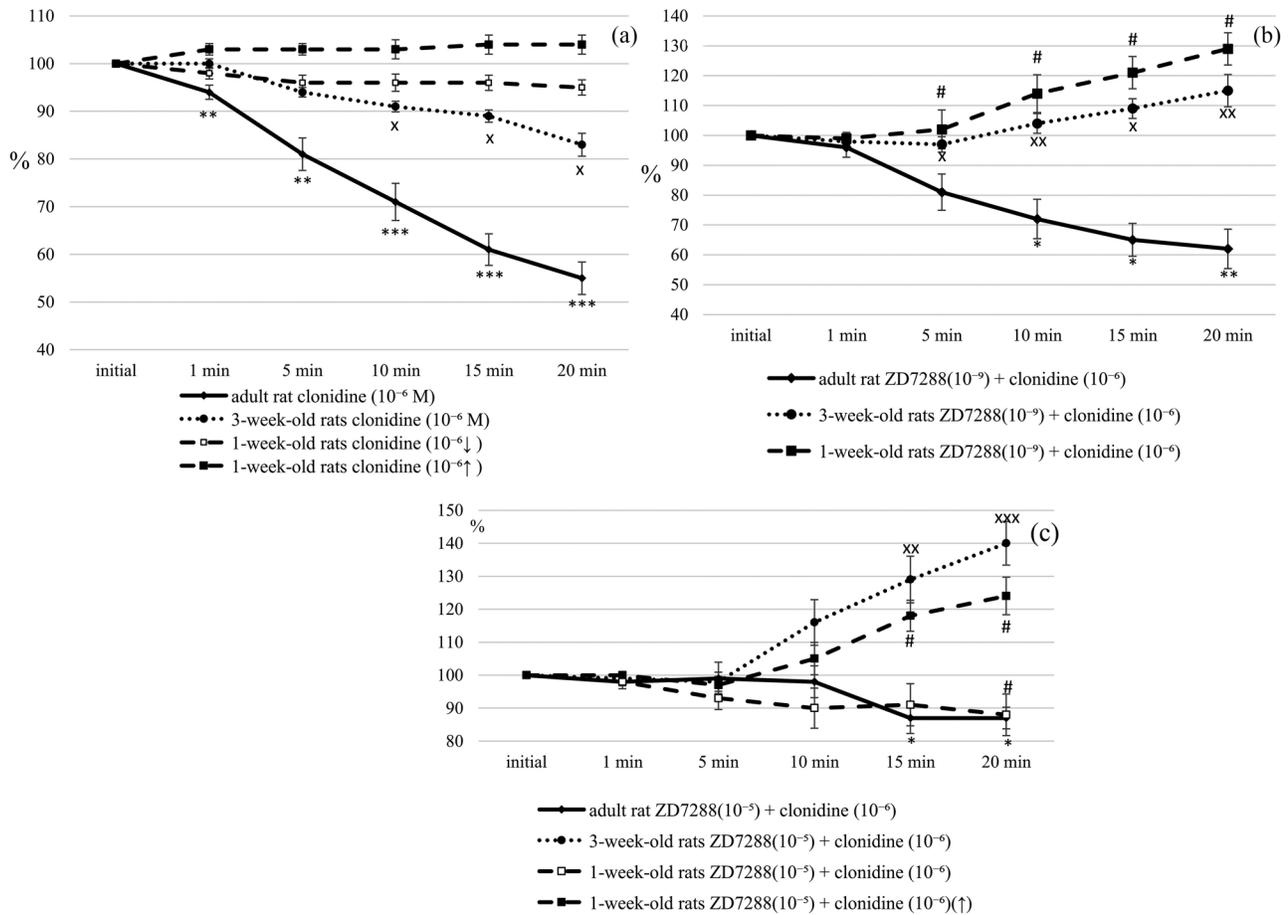
Activation of  $\alpha_2$ -ARs against the background of preliminary  $I_f$  blockade with ZD7288 ( $10^{-9}$  M) altered CF



**Fig. 1.** Effect of  $\alpha_2$ -adrenoreceptor stimulation with clonidine hydrochloride ( $10^{-6}$  M) alone (a) and against the background of  $I_f$  blockade with ZD7288 at concentrations of  $10^{-9}$  M (b) and  $10^{-5}$  M (c) on heart rate (HR) of the Langendorff heart in rats of different ages. Ordinate: HR (%), abscissa: experiment time (min). Significant differences vs. baseline values: \*X#— $p < 0.05$ ; \*\*XX— $p < 0.01$ . \*—Adult rats; X—3-week-old rat pups; #—1-week-old rat pups.



**Fig. 2.** Representative recordings of the Langendorff heart ECG in 3-week-old (a) and 1-week-old (b) rats in control and upon stimulation of  $\alpha_2$ -adrenergic receptors with clonidine hydrochloride ( $10^{-6}$  M) against the background of  $I_f$  blockade with ZD7288 ( $10^{-9}$  M).



**Fig. 3.** Effect of  $\alpha_2$ -adrenoreceptor stimulation with clonidine hydrochloride ( $10^{-6}$  M) alone (a) and against the background of  $I_f$  blockade with ZD7288 at concentrations of  $10^{-9}$  M (b) and  $10^{-5}$  M (c) on the coronary flow (CF) in the Langendorff heart in rats of different ages. Ordinate: CF (%), abscissa: experiment time (min). Significant differences vs. baseline values: \* $X^{\#}$ — $p < 0.05$ ; \*\* $XX$ — $p < 0.01$ . \*—Adult rats;  $X$ —3-week-old rat pups;  $\#$ —1-week-old rat pups.

in 1-week-old rat pups from  $0.5 \pm 0.1$  to  $0.6 \pm 0.1$  mL/min by the 5th min of observation. By the 20th min, CF increased to  $0.8 \pm 0.1$  mL/min ( $n = 7$ ,  $p < 0.01$ ) (Fig. 3b). In 3-week-old rat pups, CF increased from  $1.3 \pm 0.1$  to  $1.4 \pm 0.1$  mL/min by the 10th min. By the 20th min of the experiment, CF increased to  $1.5 \pm 0.1$  mL/min ( $n = 7$ ,  $p < 0.05$ ) (Fig. 3b). The overall change in CF amounted to 15%. In adult animals, CF decreased by 38% ( $n = 8$ ,  $p < 0.05$ ).

Activation of  $\alpha_2$ -ARs against the background of preliminary  $I_f$  blockade with ZD7288 at a higher concentration ( $10^{-5}$  M) exerted a bidirectional effect on CF in 1-day-old rat pups ( $n = 7$ ). In some of them, the baseline CF value was  $0.8 \pm 0.1$  mL/min, while by the 10th min of observation, it increased to  $0.9 \pm 0.1$  mL/min. Then, by the 20th min, it further increased by 24% vs. baseline values and reached  $1.1 \pm 0.1$  mL/min ( $n = 3$ ,  $p < 0.05$ ). In another subset

of rats, the baseline CF value was  $1.5 \pm 0.1$  mL/min, while by the 5th minute of the experiment, it decreased to  $1.4 \pm 0.1$  mL/min. Subsequently, CF was  $1.4 \pm 0.1$  mL/min by the 15th min, and changed to  $1.3 \pm 0.1$  mL/min by the 20th min ( $n = 4$ ,  $p < 0.05$ ) (Fig. 3c). The decrease amounted to 12% vs. baseline values. In 3-week-old rats, CF increased from  $1.5 \pm 0.1$  to  $1.9 \pm 0.1$  mL/min ( $p < 0.01$ ) by the 15th min of observation. Then, by the 20th min, it increased to  $2.1 \pm 0.1$  mL/min ( $n = 6$ ,  $p < 0.01$ ) (Fig. 3c). Overall, CF increased by 40% vs. baseline values. In adult animals, CF decreased by 17% ( $n = 8$ ,  $p < 0.05$ ).

## DISCUSSION

Our study on the modeling influence of  $\alpha_2$ -AR stimulation against the background of  $I_f$  blockade revealed changes in HR and CF dynamics during the

stages of adrenergic innervation formation in the isolated rat heart. In 3-week-old rat pups, the vasoconstriction of coronary vessels, observed during  $\alpha_2$ -AR stimulation, changed to vasodilation upon  $\alpha_2$ -AR stimulation with clonidine hydrochloride against the background of  $I_f$  blockade by ZD7288 ( $10^{-9}$  M,  $10^{-5}$  M), with the most pronounced effect being observed using the blocker at a higher concentration. Apparently, the formation of adrenergic fibers and receptors during this age period is associated with active regulation of the coronary bed and the involvement of  $\alpha_2$ -ARs located in both vascular smooth muscle and endothelial cells [31]. In 1-week-old rat pups,  $\alpha_2$ -AR activation caused non-significant bidirectional changes in CF. Stimulation of  $\alpha_2$ -ARs against the background of  $I_f$  blockade (ZD7288,  $10^{-9}$  M) led to a significant increase in CF, while against the background of  $I_f$  blockade at a higher blocker concentration (ZD7288,  $10^{-5}$  M), a bidirectional dynamic of CF changes was observed. However, the vasodilatory effect was more pronounced than the vasoconstrictor effect. Thus,  $\alpha_2$ -adrenergic regulation of CF in the isolated rat heart depends on the maturity level of cardiac sympathetic innervation. The maximum influence of  $\alpha_2$ -AR activation on CF was observed in adult animals, the smallest—in newborn (1-week-old) rat pups. Meanwhile, once  $I_f$  is blocked, the effect of  $\alpha_2$ -AR stimulation in newborn and 3-week-old rat pups alters the dynamics and severity of CF changes. Probably, the regulation of the coronary vascular lumen in animals of this age requires the involvement of not only  $\alpha_2$ -ARs but also hyperpolarization-activated currents, although their presence in coronary arteries is questionable, as currently there are data on the presence of HCN channels only in smooth muscle cells of lymphatic vessels and the portal vein [32].

The  $\alpha_2$ -AR agonist causes bidirectional changes in HR in 1-week-old and adult rats, and only in 3-week-old rat pups, a decrease in contraction rate was observed. Stimulation of  $\alpha_2$ -ARs against the background of  $I_f$  blockade (ZD7288,  $10^{-9}$  M) does not change the HR dynamics in 3-week-old rats and abolishes the bidirectional HR changes in adult animals, causing a reduction in HR, while in newborn rat pups, it causes an increase in HR. Since the 3-week age is characterized by a maximum HR throughout postnatal development [24], the modeling influence of  $\alpha_2$ -ARs through  $I_f$  is also aimed only at reducing HR

and may be associated with the preliminary  $I_f$  blockade that caused bradycardia. The absence of sympatho-parasympathetic interactions in newborn animals requires the involvement of various levels of  $\alpha_2$ -AR modulation to ensure the cardiac adaptation to changing conditions.

By the literature data, several mechanisms of interaction between  $\alpha_2$ -ARs and HCN channels can be assumed. Clonidine hydrochloride directly inhibits  $I_f$  [32], although our experiments demonstrate significant changes in HR and CF even against the background of HCN channel blockade. On the other hand, it has been shown that  $\alpha_2$ -AR activation via G proteins leads to the inhibition of hyperpolarization-activated currents. It is also possible that  $\alpha_2$ -ARs may be co-associated not only with the cAMP pathway but also, in some cells, with the inositol phosphate pathway. Accordingly,  $Ca^{2+}$  release upon  $\alpha_2$ -AR activation may also be associated with the activation of protein kinase C. Furthermore, the inhibitory effect of clonidine on hyperpolarization-activated currents is associated not with the cAMP level but with the activation of protein kinase C [33–34]. Against this background, the fact of the positive chronotropic effect of clonidine, which in newborn rat pups is not abolished even after HCN channel blockade, remains not entirely clear. Thus, the pathway of activation of a particular receptor apparently depends on cardiac metabolic requirements and the concentration of biologically active substances.

#### AUTHORS' CONTRIBUTION

Conceptualization and experimental design (A.M.K., N.I.Z., T.L.Z.), data collection (A.M.K., N.G.I.), data processing (A.M.K., N.G.I.), text writing and editing (A.M.K., N.I.Z., A.L.Z., T.L.Z.).

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#### ETHICS APPROVAL

Animal experiments were carried out in compliance with the NIH Guidelines for the care and use of

laboratory animals (<http://oacu.od.nih.gov/regs/index.htm>), as well as the requirements of the Basel Declaration and legal acts of the Russian Federation. Experimental protocols were approved by the Ethics Committee of Sechenov Institute of Evolutionary Physiology and Biochemistry of Kazan (Volga Region) Federal University (Minutes no. 39 of December 22, 2022).

### CONFLICT OF INTEREST

The authors of this work declare that they have no conflicts of interest.

### REFERENCES

- Zefirov TL, Ziatdinova NI, Khisamieva LI, Zefirov AL (2011) Comparative analysis of the impact of  $\alpha_1$ - and  $\alpha_2$ -adrenoceptor blockade on cardiac function in rats during postnatal ontogeny. *Bull Exp Biol Med* 151(6): 664–666. <https://doi.org/10.1007/s10517-011-1410-0>
- Zefirov TL, Ziatdinova NI, Khabibrakhmanov II, Zefirov AL (2015) Age-dependent peculiarities of the rat's heart cholinergic regulation. *Russ J Physiol* 101(2): 189–199. (In Russ)].
- Taylor EW, Leite CAC, Sartori MR, Wang T, Abe AS and Crossley DA (2014) The phylogeny and ontogeny of autonomic control of the heart and cardiorespiratory interactions in vertebrates. *J Exp Biol* 217: 690–703. <https://doi.org/10.1242/jeb.086199>
- Maltsev AV, Evdokimovskii EV, Kokoz YM (2019)  $\alpha_2$ -Adrenoceptor signaling in cardiomyocytes of spontaneously hypertensive rats starts to impair already at early age. *Biochem Biophys Res Communicat* 512: 908e913. <https://doi.org/10.1016/j.bbrc.2019.03.117>
- Robinson RB (1997) Developmental change in the voltage dependence of the pacemaker current, *if*, in rat ventricle cells. *Pflugers Arch* 433(4): 533–535. <https://doi.org/10.1007/s004240050309>
- Lipp JA, Rudolph AM (1982) Sympathetic nerve development in the rat and guinea-pig heart. *Biol Neonate* 21: 76–82. <https://doi.org/10.1159/000240497>
- Hongo M, Fujisawa S, Adachi T, Shimbo T, Shibata S, Ohba T, Ono K (2016) Age-related effects of dexmedetomidine on myocardial contraction and coronary circulation in isolated guinea pig hearts. *J Pharmacol Sci* 131(2): 118–125. <https://doi.org/10.1016/j.jphs.2016.05.002>
- Maltsev AV, Kokoz YM, Evdokimovskii EV, Pimenov OY, Reyes S, Alekseev AE (2014) Alpha-2 Adrenoceptors and Imidazoline Receptors in Cardiomyocytes Mediate Counterbalancing Effect of Agmatine on NO Synthesis and Intracellular Calcium Handling. *J Mol Cell Cardiol* 68: 66–74. <https://doi.org/10.1016/j.yjmcc.2013.12.030>
- Bao N, Tang B (2020) Organ-Protective Effects and the Underlying Mechanism of Dexmedetomidine. *Mediat Inflamm* 2020: 6136105. <https://doi.org/10.1155/2020/6136105>
- Rump LC, Riera-Knorrenschild G, Schwertfeger E, Bohmann C, Spillner G, Schollmeyer P (1995) Dopaminergic and  $\alpha$ -adrenergic control of neurotransmission in human right atrium. *J Cardiovasc Pharmacol* 26: 462–470. <https://doi.org/10.1097/00005344-199509000-00017>
- Dudek M, Knutelska J, Bednarski M, Nowiński L, Zygmunt M, Mordyl B, Głuch-Lutwin M, Kazek G, Sapa J, Pytka K (2015) Comparison of the anorectic effect and safety of the alpha2-adrenoceptor ligands guanfacine and yohimbine in rats with diet-induced. *PLoS One* 10(10): 1327–1371. <https://doi.org/10.1371/journal.pone.0141327>
- Knaus AE, Muthig V, Schickinger S, Moura E, Beetz N, Gilsbach R, Hein L (2007) Alpha2-adrenoceptor subtypes-unexpected functions for receptors and ligands derived from gene-targeted mouse models. *Neurochem Int* 51(5): 277–281. <https://doi.org/10.1016/j.neuint.2007.06.036>
- Zefirov TL, Ziyatdinova NI, Khisamieva LI, Zefirov AL (2014) Effect of  $\alpha_2$ -adrenoceptor stimulation on cardiac activity in rats. *Bull Exp Biol Med* 157(2): 154–157. <https://doi.org/10.1007/s10517-014-2523-z>
- Ziyatdinova NI, Kuptsova AM, Faskhutdinov LI, Zefirov AL, Zefirov TL (2018) Effect of  $\alpha_2$ -adrenoceptor stimulation on functional parameters of langendorff-isolated rat heart. *Bull Exp Biol Med* 165(5): 593–596. <https://doi.org/10.1007/s10517-018-4220-9>
- Kokoz YM, Evdokimovskii EV, Maltsev AV (2019) Upregulation of  $\alpha_2$ -adrenoceptor synthesis in SHR cardiomyocytes: Recompense without sense—Increased amounts, impaired commands. *Arch Biochem Biophys* 674: 108109. <https://doi.org/10.1016/j.abb.2019.108109>
- Yoshikawa Y, Hirata N, Kawaguchi R, Tokinaga Y, Yamakage M (2018) Dexmedetomidine maintains its direct cardioprotective effect against ischemia/reperfusion injury in hypertensive hypertrophied myocardium. *Anesthesia and Analgesia* 126(2): 443–452. <https://doi.org/10.1213/ANE.0000000000002452>
- DiFrancesco D (2010) The role of the funny current

- in pacemaker activity. *Circ Res* 106(3): 434–446.  
<https://doi.org/10.1161/CIRCRESAHA.109.208041>
18. Stillitano F, Lonardo G, Zicha S, Varro A, Cerbai E, Mugelli A, and Nattel S (2008) Molecular basis of funny current (If) in normal and failing human heart. *J Mol Cell Cardiol* 45: 289–299.  
<https://doi.org/10.1016/j.yjmcc.2008.04.013>
  19. Sartiani L, Mannaioni G, Masi A, Romanelli MN, Cerbai E (2017) The Hyperpolarization-Activated Cyclic Nucleotide-Gated Channels: from Biophysics to Pharmacology of a Unique Family of Ion Channels. *Pharmacol Rev* 69: 354–395.  
<https://doi.org/10.1124/pr.117.014035>
  20. Romanelli MN, Sartiani L, Masi A, Mannaioni G, Manetti D, Mugelli A, Cerbai E (2016) HCN Channels Modulators: The Need for Selectivity. *Current Top Med Chem* 16: 1764–1791.  
<https://doi.org/10.2174/1568026616999160315130832>
  21. Schweizer PA, Yampolsky P, Malik R, Thomas D, Zehelein J, Katus HA, Koenen M (2009) Transcription profiling of HCN-channel isoforms throughout mouse cardiac development. *Basic Res Cardiol* 104: 621–629.  
<https://doi.org/10.1007/s00395-009-0031-5>
  22. Mackiewicz U, Gerges JY, Chu S, Duda M, Dobrzynski H, Lewartowski B, Mączewski M (2014) Ivabradine protects against ventricular arrhythmias in acute myocardial infarction in the rat. *J Cell Physiol* 229: 813–823.  
<https://doi.org/10.1002/jcp.24507>
  23. Yampolsky P, Nauck S, Koenen M, Mosqueira M, Witzemberger M, Brueh C, Schwoerer AP, Ehmke H, Thomas D, Katus HA, Schweizer PA, Geschwill P, Seyler C, Fink T, Kruska M, Fink RHA, Draguhn A (2019) Augmentation of myocardial If dysregulates calcium homeostasis and causes adverse cardiac remodeling. *Nature Commun* 10: 3295.  
<https://doi.org/10.1038/s41467-019-11261-2>
  24. Zefirov TL, Ziyatdinova NI, Zefirov AL (2003) Effects of blockade of hyperpolarization-activated ion currents (Ih) on autonomic control of the heart in rats: Age-related peculiarities. *Neurophysiology* 35(6): 415–421.  
<https://doi.org/10.1023/B:NEPH.0000024602.05250.f1>
  25. Ziyatdinova N, Kuptsova A, Sungatullina M, Galieva A, Zefirov T (2019) Comparative analysis of the influence of If blockade on newborn and adult rats Langendorff-isolated heart. *Eur J Clin Inv* 49: 144.
  26. Kuptsova AM, Ziyatdinova NI, Zefirov TL (2019) The Effect of the Pacemaker Current Blockade on the Rat Heart during the Formation of the Adrenergic Innervation. *Russ J Physiol* 105(10): 1294–1304. (In Russ).  
<https://doi.org/10.1134/S0869813919100042>
  27. Zefirov TL, Gibina AE, Sergejeva AM, Ziyatdinova NI, Zefirov AL (2007) Age-Related Peculiarities of Contractile Activity of Rat Myocardium during Blockade of Hyperpolarization-Activated Currents. *Bull Exp Biol Med* 144(3): 273–275.  
<https://doi.org/10.1007/s10517-007-0308-3>
  28. Bucchi A, Baruscotti M, Robinson RB, DiFrancesco D (2007) Modulation of rate by autonomic agonists in SAN cells involves changes in diastolic depolarization and the pacemaker current. *J Mol Cell Cardiol* 43: 39–48.  
<https://doi.org/10.1016/j.yjmcc.2007.04.017>
  29. Qu J, Cohen IS, Robinson RB (2000) Sympathetic innervation alters activation of pacemaker current (If) in rat ventricles. *J Physiol* 526: 561–569.  
<https://doi.org/10.1111/j.1469-7793.2000.t01-1-00561.x>
  30. Robinson RB (1996) Autonomic receptor-effector coupling during postnatal development. *Cardiovasc Res* 31: 68–76.
  31. Kozlovsky VI (2010) The role of the endothelium in vasodilation mediated by various subtypes of adrenergic receptors. *J GrSMU* 1: 32–35. (In Russ).
  32. Benzoni P, Bertoli G, Giannetti F, Piantoni C, Milanese R, Pecchiari M, Barbuti A, Baruscotti M, Bucchi A (2021) The funny current: Even funnier than 40 years ago. Unconventional expression and roles of HCN/f channels all over the body. *Progr Biophys Mol Biol* 166: 189e204.  
<https://doi.org/10.1016/j.pbiomolbio.2021.08.007>
  33. Knaus A, Zong X, Beetz N, Jahns R, Lohse MJ, Biel M, Hein L (2007) Direct Inhibition of Cardiac Hyperpolarization-Activated Cyclic Nucleotide-Gated Pacemaker Channels by Clonidine. *Arrhythmia/Electrophysiol* 115: 872–880.  
<https://doi.org/10.1161/CIRCULATIONAHA.106.667675>
  34. Ying-cong Y, Qing-tao M, Xia P, Zhong-yuan X, Xiang-dong C (2014) Dexmedetomidine produced analgesic effect via inhibition of HCN currents. *Eur J Pharmacol* 740: 560–564.  
<https://doi.org/10.1016/j.ejphar.2014.06.031>

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