A SIMPLE METHOD FOR ISOLATION OF SOME *BACILLUS* STRAINS WITH AN EXPRESSED ANTI-CANCER ACTIVITY

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ABSTRACT:

There is now increasing evidence that probiotic bacteria can provide health benefits to humans. In many areas of medicine (gastroenterology, urology, allergology, oncology and others), these sanative microorganisms may be considered as possible and viable alternatives applicable to patient care. Particularly, we have found that oral administration of *Bacillus oligonitrophilus* KU-1 cells can be used for treatment and prevention of some tumors. Here we present a simple method for isolation of bacteria with anticancer properties from soil.

Keywords: probiotics, cancer, Bacillus oligonitrophilus, soil.

RESUMEN:

Está aumentando la evidencia de que hay bacterias probióticas que pueden proporcionar beneficios saludables a los seres humanos. En muchas áreas de la medicina (gastroenterología, urología, alergología, oncología y otras), estos microorganismos pueden considerarse como alternativas posibles y viables aplicables al cuidado del paciente. Particularmente, nosotros hemos encontrado que la administración oral de células KU-1 *Bacillus oligonitrophilus* puede ser utilizada para el tratamiento y la prevención de algunos tumores. Aquí presentamos un método simple para aislamiento de suelos, de bacterias con características anticáncer.

Palabras Clave: Probióticos, cáncer, Bacillus oligonitrophilus, suelo.

INTRODUCTION

Health benefits of probiotics have been well documented^{1, 2}. In the last decade, many investigators have studied the therapeutic and prophylactic effects of some probiotic bacteria and found that oral administration of such microorganisms might modulate pattern of cancer and other diseases³⁻¹⁴. Microorganisms most commonly used as probiotics are lactic bacteria (for example, lactobacilli and bifidobacteria). However, it has been showed lately that other non-dairy microorganisms might also be used as probiotics¹⁵⁻¹⁸.

In 2005, we reported that soil bacterium *Bacillus oligonitrophilus* KU-1 was used successfully as a monotherapeutic drug in cancer patients¹⁹. The theoretic background for use of silicate-degrading bacteria like B. oligonitrophilus KU-1 in medicine in was reported previously by Voronkov and co-workers²⁰⁻²² and discussed in our recent publication²³.

Here we present a simple technique for isolation from soil of some Bacillus strains with anticancer activity.

MATERIALS AND METHODS

Media.

All chemicals are reagent grade unless otherwise noted. Contents of the used media are presented in Table 1. The mixtures are brought to 1,000 ml with distilled H_2O and autoclaved.

Table 1. Content of incrobiological media used for isolation and cultivation of son baching					
Medium	Ingredients (g/L) and pH				
Giss	Peptone 3, 65; agar 4.11; mannitol – 2.85; Na ₂ HPO ₄ 0.77; NaCl 3.55; aqua blue 0.03; aurin				
medium	0.03; pH 7.4±0.2.				
Ploscirewi	Peptone 16; agar 8.75; salts of bile acids 8.1; lactose 7.6; sodium hydrocitrate disubstituted				
medium	8.82; neutral red 0.04; brilliant green 0.02; Na ₂ HPO ₄ 2.25; sodium thiosulfate 6.86; metallic				
	iodine (I ₂) 0.12; Na ₂ CO ₃ 1.42; pH 7.1-7.3.				
Alexandrov	Na ₂ HPO ₄ 10; (NH ₄) ₂ SO ₄ 2; MgSO ₄ 0.5; SiO ₂ 0.15; CaCO ₃ 0.05; orthoclase 0.5; pH 8.0.				
medium					
Nutrient	Peptone 5; meat extract 3; agar 15; $MnSO_4xH_2O$ 0.01; pH 7.0.				
agar					

Table 1. Content of microbiological media used for isolation and cultivation of soil bacilli

Extraction from soil and identification of bacteria. Bacteria are extracted from soil samples taken from Samara Luka (Samara Region, Russia) by placing 500 mg of soil into a sterile Erlenmeyer flask with 2 mL of sterile distilled water. The flask is shaken at 200 rpm for 5 minutes and 20 μ L of the slurry is serially inoculated to various growth media (Giss, Ploscirewi, Alexandrov media).

After incubation in Giss medium with mannitol at 22 ⁰C for 24 h, single dark blue bacterial colonies should be scrapped by using a microbiological loop and cells should be drifted to Giss media with various carbohydrate supplements. In Giss medium with glucose, bacilli with anticancer activity are permanently dark blue. In Giss media with saccharose and lactose, these bacteria are usually dark blue. In Giss media with maltose, anticancer bacteria are rarely dark blue.

Then, the selected bacteria should be tested in Ploscirewi medium, which can be used for selection of Zn group (*Bacillus oligonitrophilus* KU-2) as well as other groups with possible anti-cancer activity. These groups are resistant to various disturbing factors (diverse salts, etc) in this medium.

Further, Alexandrov medium with glucose (at 22 0 C for 24-48 h without aeration, pH=8.0) may be used for selection: anticancer bacteria shows suspension growth (after 1-2 days) with bubble formation. During bacterial growth, the acidity of the medium decreases usually to a pH value of 4.0-4.5.

Antimicrobial sensitivity.

Finally, typing with antibiotics on the nutrient agar can also be used for selection of anticancer bacteria.

Antibiotic	Diameter of inhibition zone (mm)			
	Bacillus oligonitrophilus		Bacillus mucilaginosus	
(a dose is indicated	Z _n (KU-2)	K _x -53 (KU-1)	D-2 (KU-6)	D _a (KU-5)
in brackets)				
Ampicillin (30 ì g)	15	15	16	17
Doxycyline (10 ì g)	11	12	10	11
Kanamycin (30 ì g)	16	17	16	16
Carbenicillin (25 ì g)	9	16	18	6
Chloramphenicol (30 ì g)	20	23	20	18
Monomycin (30 ì g)	0	0	0	0
Neomycin (30 ì g)	16	16	15	17
Oleandomycin (15 ì g)	0	0	0	0
Polymixin (300 IU)	12	12	11	13
Rifampicin (5 ì g)	0	0	0	0
Rystomycin (30 ì g)	0	0	0	0
Fuzidin (10ìg)	0	0	0	0
Streptomycin (30 ì g)	12	13	14	6
Cefalexin (30 ì g)	0	0	0	0
Erythromycin (15 ì g)	6	7	9	6

Table 2. Typing with antibiotics in Bacillus strains on nutrient agar

Note: 0 – total tolerance, sensibility – diameter of inhibition zone more than 15 mm.

Table 2 presents data on typing with antibiotics.

RESULTS AND DISCUSSION

Using the above-mentioned techniques, we have isolated four bacterial strains: Zn (KU-2), Kx-53 (KU-1), D-2 (KU-6), Da (KU-5). The first two strains belong to B. oligonitrophilus while the second two are B. mucilaginosus (according to 24). Anticancer activity of B. oligonitrophilus KU-1 has been tested with success in cancer patients¹⁹. Broad clinical trials are warranted to test the other strains.

It should be noted here that the use Giss media reveals a maximal ability of these bacteria to synthesize organic acids. This is of great significance because organic acids (succinic acid, lactic acid, etc) may inhibit saprogenous bacteria in the gastrointestinal tract reducing formation of carcinogens²⁵⁻²⁶.

The subsequent selection in Ploscirewi medium makes possible to identify bacteria (normal growth, weak growth or absence of growth) that are resistant to severe conditions. This medium is considered to be unsuitable for growth of many bacteria because it contains metallic iodine, which is toxic to the bacteria. Moreover, bacteria grown in Ploscirewi medium should be tolerant to bile acids. This is very important for survival of bacteria in the human gastrointestinal tract and for their prolonged and efficient action there. Growth in Alexandrov medium and antibiotic-resistance tests is the final stages of selection.

By our experience, bacteria with anticancer features may be isolated with increased probability from soils in regions where geotectonic ruptures take place. It is very likely to isolate these bacteria from Alpine areas of Pamir, Cordilleras and Andes. However, our suggestions should be checked experimentally.

We hope that the presented material can be used for isolation of bacilli with anticancer activity and for further human benefit maintaining the balance of the gut microflora and favoring the equipoise in healthy and diseased individuals. 1 Fuller R. Probiotics in human medicine. Gut 1991; 32: 439-442.

2 Bourlioux P., Bouley C, Ashwell M. New aspects of the functionalities of probiotics. Forum Nutr 2003; 56: 355-356.

3 Reddy B, Rivenson A. Inhibitory effect of *Bifidobacterium longum* on colon, mammary, and liver carcinogenesis induced by 2-amino-3-methylimidazo(4-5)quinoline, a food mutagen. Cancer Res 1993; 53: 3914-3918.

4 Aso Y, Akaza H, Kotake T, Tsukamoto T, Imai K, Naito S. Preventive effect of a *Lactobacillus casei* preparation on the recurrence of superficial bladder cancer in a double-blind trial. The BLP Study Group. Eur Urol 1995; 27: 104-109.

5 Hosoda M, Hashimoto H, He F, Morita H, Hosono A. Effect of administration of milk fermented with *Lactobacillus acidophilus* LA-2 on fecal mutagenicity and microflora in the human intestine. J Dairy Sci 1996; 79: 745-749.

6 Perdigon G, Valdez JC, Rachid M. Antitumour activity of yogurt: study of possible immune mechanisms. J Dairy Res 1998; 65: 129-138.

7 Brady LJ, Daniel G, Busta D. The role of probiotic cultures in the prevention of colon cancer. J Nutr 2000; 130: 410S-444S.

8 Rachid MM, Gobbato NM, Valdez JC. Effect of yogurt on the inhibition of an intestinal carcinoma by increasing cellular apoptosis. Int J Immunopathol Pharmacol 2002; 15; 209-216.

9 Philpott M, Ferguson LR. Immunonutrition and cancer. Mutation Res 2004; 551: 29-42.

10 Rowland I. Probiotics and colorectal cancer risk. Brit J Nutr 2004; 91: 805-807.

11 Vorob'ev AA, Gershanovich ML, Petrov LN. Prerequisites and perspectives for use of probiotics in complex therapy of cancer. Voprosy Onkologii 2004; 50: 361-365 (In Russian).

12 de Moreno de LeBlanc A, Perdigon G. Reduction of beta-glucuronidase and nitroreductase activity by yoghurt in a murine colon cancer model. Biocell 2005; 29: 15-24.

13 Maksimov IK, Kalinin AV, Rukavitsyn OA, Minushkin ON, Ardamskaia MD. The condition of intestinal microflora in patients with hematologic tumors receiving polychemotherapy. Klinitcheskaya Meditzina (Mosk) 2005; 83: 48-53. (In Russian).

14 Mego M, Ebringer L, Drgona L, Mardiak J, Trupl J, Greksak R, Nemova I, Oravcova E, Zajac V, Koza I. Prevention of febrile neutropenia in cancer patients by probiotic strain *Enterococcus faecium* M-74. Pilot study phase I. Neoplasma 2005; 52: 159-164.

15 Urdaci MC, Bressollier P, Pinchuk I. *Bacillus clausii* probiotic strains: antimicrobial and immunomodulatory activities. J Clin Gastroenterol 2004; 38(6 Suppl): S86-90.

16 Nista EC, Candelli M, Cremonini F, Cazzato IA, Zocco MA, Franceschi F, Cammarota G, Gasbarrini G, Gasbarrini A. *Bacillus clausii* therapy to reduce side-effects of anti-Helicobacter pylori treatment: randomized, double-blind, placebo controlled trial. Aliment Pharmacol Ther 2004; 20: 1181-1188.

17 Brown AC, Shovic A, Ibrahim SA, Holck P, Huang A. A non-dairy probiotic's (poi) influence on changing the gastrointestinal tract's microflora environment. Altern Ther Health Med 2005; 11: 58-64.

18 Ciprandi G, Vizzaccaro A, Cirillo I, Tosca MA. Bacillus clausii exerts immuno-modulatory activity in allergic subjects: a pilot study. Allergy Immunol (Paris) 2005; 37: 129-134.

19 Malkov SV, Markelov VV, Polozov GY, Sobchuk LI, Zakharova NG, Barabanschikov BI, Kozhevnikov AY, Vaphin RA, Trushin MV. Antitumor features of *Bacillus oligonitrophilus* KU-1 strain. J Microbiol Immunol Infect 2005; 38: 96-104.

20 Voronkov MG, Grigalinovich GA, Zelchan GI. Inhibitor action of some silicon compounds on the growth of malignant tumor cells. Doklady Akademii Nauk SSSR 1971; 200: 967-969. (In Russian).

21 Voronkov MG, Skorobogatova VI, Vugmeister EK, Makarskii VV. Silicon in nucleic acids. Doklady Akademii Nauk SSSR 1975; 220: 723-725. (In Russian).

22 Voronkov MG, Kuznetzov IG. Silicon in living organism, Novosibirsk: Nauka, 1984. (In Russian).

23. Malkov SV, Markelov VV, Barabanschikov BI, Trushin MV, Marotta F. Genome rejuvenation and its practical applications. The Biomedical Scientist 2006; 50: 45-47.

24. Krasilnikov NA, eds. Opredelitel bakterii i actinomycetov (Manual for identification of bacteria and actinomycete). Moscow-Leningrad: USSR Academy of Sciences Press; 1949.

25 Reid G, McGroarty JA, Angotti R, Cook RL. Lactobacillus inhibitor production against *Escherichia coli* and coaggregation ability with uropathogens. Canad J Microbiol 1988; 34: 344-351.

26. Zhang XB, Ohta Y. Antimutagenicity and binding of lactic acid bacteria from Chinese cheese

to mutagenic pyrolyzates. J Dairy Sci 1990; 73: 2702-2710.

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This paper reports a method for isolation of some *Bacillus* strains that might have probiotic properties. There are some publications showing that these bacterias might have anticancer activity. The reported method may be useful for obtaining large amounts of these bacteria that allow the realisation of controlled clinical trials to confirm if they are beneficial for treatment or prevention of cancer.

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Cancer is doubtlessly one of the most difficult diseases to treat. Multiple mechanisms of tumor expression and resistance to medications complicate the search for a cure. New alternatives are always welcomed. In this article, the authors present a simple method for isolation of bacteria that have shown antitumor properties and have been studied in cancer patients. Further studies are needed, however, and the publication of the method of isolation may be an important first step in that direction.

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