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# Hydrogen Sulfide in Regulation of Frog Myocardium Contractility<sup>1</sup>

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Abstract—Hydrogen sulfide (H<sub>2</sub>S) is an endogenously synthesized gaseous molecule which, along with nitric oxide and carbon monoxide, induces a number of effects in cardiovascular system under normal and pathological conditions. In the present work, the effects and underlying mechanisms of the H<sub>2</sub>S donor sodium hydrosulfide (NaHS) on the isometric force of frog myocardium contraction have been studied. NaHS at the concentration of 100 µM induced negative inotropic effect and reduced the maximum velocity of the contraction and relaxation of the isolated ventricle strips. The substrate of  $H_2S$  synthesis, L-cysteine (200  $\mu$ M and 1 mM), induced the same effect, while the inhibitors of cystathionin- $\gamma$ -Jyase, the H<sub>2</sub>S-producing enzyme in heart,  $\beta$ -cyanoalanine (500  $\mu$ M) and propargylglycine (500  $\mu$ M), increased the amplitude of contraction. Inhibition of cystathionin- $\gamma$ -lyase by  $\beta$ -cyanoalanine prevented the negative inotropic effect of L-cysteine. After the inhibition of adenylate cyclase by MDL-12,330A (3  $\mu$ M) or phosphodiesterases by IBMX (200  $\mu$ M), the effect of NaHS was less than that in the control. In the presence of membrane-penetrating analogous of cAMP, 8BrcAMP (100  $\mu$ M) and pCPT-cAMP (100  $\mu$ M), the negative inotropic effect of NaHS was completely retained. The effect of NaHS significantly decreased after preliminary application of the NO donor, SNAP  $(10 \,\mu\text{M})$ , and did not change after the inhibition of NO syntheses by L-NAME (100  $\mu\text{M}$ ). The results suggest the possibility of endogenous synthesis of H<sub>2</sub>S in frog myocardium and regulation of its contractility by the activation of phosphodiesterases hydrolyzing cAMP, which leads to a decrease in the activation of cAMPdependent protein kinases and phosphorylation of voltage-dependent L-type Ca channels. As a result, the reduction of calcium entry into cardiomyocytes decreases the contractility of frog myocardium.

*Keywords*: hydrogen sulfide, myocardial contractility, adenylate cyclase, nitric oxide **DOI:** 10.1134/S1990747812030117

Hydrogen sulfide  $(H_2S)$  is known as a toxic gas [1]; however, the more emerging evidence shows that  $H_2S$ is synthesized endogenously and has various physiological effects on the cardiovascular and nervous systems, as well as on gastrointestinal tract [2-7]. It is supposed that  $H_2S$  is an endogenous "gasotransmitter", like two other physiologically active gasses: nitric oxide (NO) and carbon monoxide (CO) [8, 9]. In the cardiovascular system,  $H_2S$  is synthesized from L-cysteine by cystathionin-y-lyase and 3-mercaptosulfotransferase [5, 10, 11]. It induces vasodilation and regulates proliferation, apoptosis, and angiogenesis. In addition, H<sub>2</sub>S has a cardioprotective effect against injuries induced by ischemia-reperfusion [5, 10, 12]. It is interesting that H<sub>2</sub>S influences the vascular tone of all classes of vertebrates (fish, amphibian, reptiles), including vasoconstriction and vasodilation, which indicates the phylogenetic antiquity of H<sub>2</sub>S as a gasotransmitter and universality of its action [13, 14].

It was shown in the intact rat heart and isolated cardiomyocytes that  $H_2S$  reduced the duration of action potential and had a negative inotropic effect [10, 15, 16]. The mechanisms of H<sub>2</sub>S action in mammalian myocardium include the adenylate cyclase system, ATP-dependent K channels, and voltage-dependent L-type Ca channels, according to data from various sources and depending on the animal species [11, 15–17]. The regulation of cAMP synthesis by H<sub>2</sub>S may play a critical role in cardioprotection, as there is a substantial decrease in H<sub>2</sub>S production in different models of ischemia and hyperstimulation of β-adrenoceptors [10, 12]. In the frog myocardium, the hydrogen sulfide donor NaHS had a dose-dependent negative inotropic effect [18], but the mechanisms of H<sub>2</sub>S action were not revealed.

The goal of this work was to study the effects of exogenous and endogenous  $H_2S$  on frog myocardial contractility and to elucidate the plausible molecular mechanisms of its action.

# MATERIALS AND METHODS

The experiments on contractility recording were performed on the strips of frog myocardium in a Pow-

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erLab device using an isometric transducer with a sensitivity of 0–50 g (AD Instrument, Australia, and Biopac, USA). A ventricular myocardium strip of 4-6 mm in length and 1 mm in diameter was submerged vertically in the chamber with the working volume of 20 ml and perfused with Ringer's solution for cold-blooded animals containing (mM): 118.0 NaCl, 2.5 KCl, 1.8 CaCl<sub>2</sub>, 10 Trizma (pH 7.3–7.4, 20°C). The preparation was stimulated by electric pulses at 0.1 Hz via two silver electrodes. The signals were processed with the Chart software; the force of contraction was determined in grams. The contraction amplitude and the maximum rates of contraction and relaxation were estimated. Statistical analysis was performed using the standard methods; statistical significance of the differences was determined by Student's *t*-criterion.

Sodium hydrosulfide (NaHS) was used as a donor of H<sub>2</sub>S; it dissociates into Na<sup>+</sup> and HS<sup>-</sup> in aqueous solution; then  $HS^-$  is bound to  $H^+$  forming  $H_2S$ . In the neutral solution, one third of NaHS is present as gaseous  $H_2S$  and two thirds are present as  $HS^-$  [1]. The following agents were used in the experiments: L-cysteine,  $\beta$ -cyanalanine, propargylglycine, IBMX (3isobutyl-1-methylxantine), 8Br-cAMP, pCPT-cAMP (sodium salt of 8-(4-chlorophenylthio)adenosine-3',5' cyclophosphate), MDL-12,330A (hydrochloride of cis-N-(2-phenylcyclopentyl)azacyclotridec-1-en-2-amine), L-NAME (N<sup>G</sup>-nitro-*L*-arginine methyl ester). SNAP (S-nitroso-N-acetylpenicillamine). Water-insoluble substances were dissolved in dimethvlsulfoxide (DMSO). DMSO concentration in the solution did not exceed 0.01%. At this concentration, DMSO in the control experiments had no substantial effect on the force of myocardial contraction. All substances were produced by Sigma (USA).

#### RESULTS

The effects of the donor, substrate and inhibitors of H<sub>2</sub>S synthesis on the force of myocardial contraction. The  $H_2S$  donor NaHS at the concentration of 100  $\mu$ M caused a reversible decrease in the amplitude of contraction of the frog heart ventricle strip to  $66 \pm 6\%$  (n = 14, p < 0.05) by the 20th min of application, which is in agreement with our previous data [18]. At the same time, NaHS reduced the maximum rates of the myocardium strip contraction and relaxation to  $57 \pm 8$  and  $53 \pm 7\%$  (n = 5, p < 0.05) compared to the control, respectively. The substrate of the  $H_2S$  synthesis, L-cysteine, by the 20th min of application caused, like the gas donor, the decrease in the amplitude of myocardial strip contraction to  $83 \pm 6\%$  (n = 6, p < 0.05) at a concentration of 200  $\mu$ M and to 58 ± 4% (n = 7, p < 0.05) at a concentration of 1 mM (Fig. 1a, b).

The possibility of endogenous gas synthesis was revealed using cystathionin- $\gamma$ -lyase inhibitors ( $\beta$ -cyanoalanine and propargylglycine) at the concentration of 500  $\mu$ M.  $\beta$ -Cyanoalanine and propargylglycine increased the force of myocardial contraction to 117 ± 6% (n = 8, p < 0.05) and 112 ± 3% (n = 8, p < 0.05), respectively, by the 15th minute of application (Fig. 1b). Thus, the endogenously synthesized H<sub>2</sub>S induced the same effects as the exogenously applied H<sub>2</sub>S, while cystathionin- $\gamma$ -lyase inhibitors exerted an opposite effect. Under the conditions of cystathionin- $\gamma$ -lyase inhibition by  $\beta$ -cyanoalanine, the application of *L*-cysteine (200  $\mu$ M or 1 mM) did not lead to the decrease in the contraction amplitude.

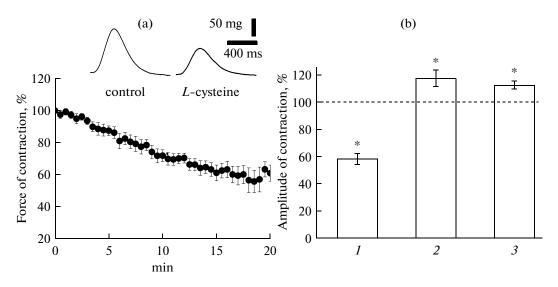
Adenylate cyclase system and H<sub>2</sub>S effects in myocardium. In the nervous system and cardiomyocytes of warm-blooded animals, H<sub>2</sub>S effects are probably mediated by the changes in cAMP level [11, 19]; therefore, the effect of the gas was studied under the increase and decrease in the level of this cyclic nucleotide. Adenylate cyclase inhibition by MDL-12,330A (3  $\mu$ M) reduced the force of contraction to 81 ± 3% (*n* = 11, *p* < 0.05) by the 25–30th minute of application (Fig. 2), which is probably associated with the decrease of cAMP concentration in cardiomyocytes. Under these conditions, the effect of H<sub>2</sub>S was less pronounced than in the control: 87 ± 2% (*n* = 13, *p* < 0.05) (Fig. 2).

cAMP concentration was increased by using the membrane penetrating analogs: 8Br-cAMP and pCPT-cAMP, at a concentration of 100  $\mu$ M. The application of 8Br-cAMP or pCPT-cAMP did not cause significant changes in the amplitude of contraction. By the 20th minute of pCPT-cAMP or 8Br-cAMP action, the force of myocardium strip contraction was 110 ± 11% (n = 5, p > 0.05) or 101 ± 5% (n = 5, p > 0.05), respectively, compared to the control values. The addition of NaHS reduced the strip contractility to 70 ± 6% (n = 5, p < 0.05) in the presence of 8Br-cAMP and to 57 ± 11% (n = 5, p < 0.05) in case of pCPT-cAMP (Fig. 2). Thus, the effect of NaHS under the influence of cAMP analogs was maintained to the same extent as in the control conditions.

Phosphodiesterase inhibition is another way to increase the level of cAMP in the cells. IBMX, a non-specific blocker of phosphodiesterases of the cyclic nucleotides, was used in our experiments at the concentration of 200  $\mu$ M. IBMX increased the force of contraction by 120 ± 4% (n = 7, p < 0.05) by the 8th minute of application, probably due to the accumulation of cAMP in cardiomyocytes (Fig. 2). At the enhanced level of endogenous cyclic nucleotides, NaHS reduced the force of myocardium strip contraction to  $82 \pm 4\%$  (n = 5), and this effect was less than the effect of H<sub>2</sub>S donor in the control (p < 0.05) (Fig. 2).

Thus, the effect of  $H_2S$  was partially eliminated under the inhibition of adenylate cyclase or phosphodiesterases but maintained under the action of cAMP analogs.

The nitric oxide system and the effects of hydrogen sulfide. It is known that NO is an important regulator of the force of myocardial contraction in frog [20]. Based on the data on interaction between the systems of gaseous mediators, we analyzed the effects of NaHS



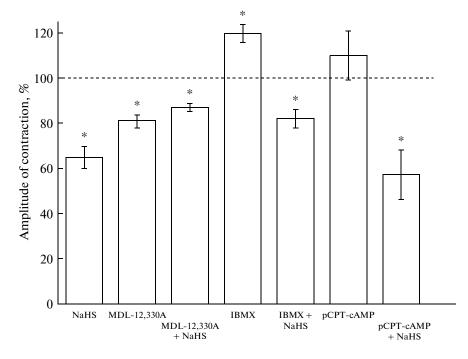
**Fig. 1.** The effects of the substrate and inhibitors of hydrogen sulfide synthesis on myocardial contractility. (a) The dynamics of contraction force under the action of the H<sub>2</sub>S synthesis substrate, *L*-cysteine, at a concentration of 1 mM. On top, the original recordings of contraction in the control and by the 20th minute of *L*-cysteine application are shown. (b) The effects of 1 mM *L*-cysteine (*1*) and cystathionin-γ-lyase blockers: 500 µM β-cyanalanine (*2*) and 500 µM propargylglycine (*3*) on the force of myocardial contraction. \**p* < 0.05.

under the conditions of NO synthesis inhibition and increase of its endogenous concentration. The application of L-NAME, the blocker of different forms of NO synthases (100  $\mu$ M), resulted in a slight increase in the amplitude of contractions. By the 15th min of L-NAME application, the force of contraction was  $115 \pm 4\%$  (n = 7, p < 0.05). Under the inhibition of NO synthesis, NaHS reduced the force of myocardial contraction to  $65 \pm 8\%$  (*n* = 7), which was not different from the effect of H<sub>2</sub>S in the control. Thus, the effect of H<sub>2</sub>S is not associated with the changes in NO synthesis. The NO donor (SNAP) at the concentration of 10  $\mu$ M reduced myocardial contractility to 86  $\pm$  5% (n = 10, p < 0.05). Under the influence of SNAP, the negative inotropic effect of NaHS was much less pronounced than in the control:  $89 \pm 2\%$  (n = 11, p < 0.05).

## DISCUSSION

The negative inotropic effect of exogenous and endogenous  $H_2S$ . We have shown that both exogenous and endogenously synthesized  $H_2S$  has a negative inotropic effect on the frog myocardium. The substrate of  $H_2S$  synthesis *L*-cysteine caused the same decrease in the amplitude of myocardial contractions as the  $H_2S$ donor NaHS, whereas the cystathionin- $\gamma$ -lyase inhibitors had an opposite effect: increased the amplitude of the contraction. It is known that *L*-cysteine is present in mammalian plasma in micromolar concentrations, but the enzymes of  $H_2S$  synthesis have low affinity to cysteine. Therefore,  $H_2S$  synthesis is usually enhanced by using high substrate concentrations: 1 mM and more [21–23]. In our experiments, the force of myocardial contraction decreased already at 200  $\mu$ M and was well pronounced at 1 mM of the substrate, probably due to intensification of H<sub>2</sub>S synthesis. In addition, the effect of L-cysteine was not manifested during the inhibition of cystathionin- $\gamma$ -lyase (Fig. 1). The findings demonstrate the possibility of H<sub>2</sub>S synthesis in the myocardium of cold-blooded animals by cystathionin- $\gamma$ -lyase, with L-cysteine as a substrate. The negative inotropic effect of NaHS in the myocardium was observed in warm-blooded animals [10, 11, 16] and in our previous works with frog heart [18]. There are no data on gas concentration in frog myocardium. However, H<sub>2</sub>S is synthesized in the vessels of lower vertebrates by cystathionin- $\gamma$ -lyase at concentrations comparable with the H<sub>2</sub>S concentration in rat vessels. The  $H_2S$  concentration in the plasma of trout is approximately 40 µM, i.e., higher than in mammals [13, 14]. It seems that the myocardium of frog, similar to the vertebrates of other classes, contains a H<sub>2</sub>S synthesis system, which implies its tonic effect on the myocardium.

cAMP as a factor mediating the effect of  $H_2S$  on myocardial contractility. The main factor that triggers and determines the force of myocardial contractility is intracellular concentration of calcium ions. The voltage-dependent L-type Ca channels open in response to the cardiomyocyte membrane depolarization followed by the entry of Ca<sup>2+</sup>, which induces Ca<sup>2+</sup> release from the intracellular Ca stores via ryanodine receptors and triggers the process of muscular contraction [24, 25]. In the ventricular cardiomyocytes of frogs, as a result of poor development of sarcoplasmic reticulum, the main calcium source is Ca<sup>2+</sup> entering via the Ca channels of the plasma membrane [26]. The negative inotropic effect of NaHS is probably due to the



**Fig. 2.** The effect of sodium hydrosulfide at a decreasing and increasing level of cyclic nucleotides. The amplitudes of myocardial contraction are shown under the action of NaHS (100  $\mu$ M) in the control, MDL-12,330A (3  $\mu$ M), IBMX (200  $\mu$ M), pCPT-cAMP (100  $\mu$ M) and NaHS under the action of the above reagents. During the analysis of NaHS effects, the values obtained under the action of MDL-12,330A, IBMX, and pCPT-cAMP were taken as 100%, respectively. \**p* < 0.05.

decrease in the entry of calcium ions into cardiomyocytes. In our previous studies, the NaHS effect was completely preserved under the conditions of Ca channels inhibition by nifedipine [18], which indicates the absence of direct inhibitory effect of the gas on Ca-channel subunits. The Ca current may be regulated through phosphorylation by cAMP-dependent kinases at an enhanced level of cAMP in response to the activation of  $\beta$ -adrenoceptors [27]. It is known that the Ca channel in phosphorylation state is characterized by the higher conductivity and open probability and activated at more negative values of membrane potential [25].

In our experiments, both the decrease and the increase in the endogenous level of cAMP reduced the manifestation of  $H_2S$  effect, demonstrating the involvement of adenylate cyclase system in these effects (Fig. 2). At the same time, the adenylate cyclase inhibition reduced the force of contraction, while the increase in the endogenous level of cAMP by the nonspecific phosphodiesterase inhibitor IBMX intensified the response, which confirms the involvement of cAMP-dependent pathways in contractility regulation. However, the application of membranepenetrating cAMP analogs (pCPT-cAMP or 8BrcAMP) did not result in any substantial changes in contraction intensity nor influenced the effect of NaHS (Fig. 2). The analogs used are relatively stable, though they are known to be subject to hydrolysis by Ca/calmodulin-dependent, cGMP-inhibited and cGMP-stimulated phosphodiesterases [28] expressed in frog cardiomyocytes. In addition, the absence of the effect of pCPT-cAMP or 8Br-cAMP on the force of myocardial contraction indicates that they cannot simulate the situation when the endogenous level of cAMP increases (like in the case with IBMX). It may be due to the fact that cAMP analogs activate not only protein kinase A but also protein kinase G [28, 29] and inhibit the cGMP-specific phosphodiesterase, which results in enhancement of the cGMP level [28, 30]. The activation of protein kinase G, in turn, will lead to a decrease in the Ca current amplitude [31]. In addition, an important factor of cAMP influence on contractility is its localization close to the targets, while mere increase in the total level of cAMP is insufficient for specific regulation of protein target [32]. In the ventricular cardiomvocvtes of frog. Ca current is regulated by the local increase in cAMP level close to the cytoplasmic membrane. Phosphodiesterases provide cAMP compartmentalization, preventing its diffusion along the cardiomyocyte. The location of A-kinase anchoring protein close to the Ca channels provides the local control of Ca current by cAMP-dependent processes [33].

In accordance with the published data, adenylate cyclase may be a target for NaHS. In the central nervous system, NaHS enhanced the conductance of glutamate NMDA-receptors and increased the cAMP level [19]. In rat cardiomyocytes,  $H_2S$  reduced the positive inotropic effect of isoproterenol due to the

inhibition of cAMP synthesis [11]. The involvement of the cAMP system in  $H_2S$  effects was noted also in the motor nerve ending of frog, where the effect of the gas decreased at an enhanced level of cAMP [34].

The role of nitric oxide in H<sub>2</sub>S effects. Our data show the interaction between the two systems of gaseous mediators: H<sub>2</sub>S and NO. So, after preliminary application of the NO donor (SNAP), the effect of NaHS was less pronounced than in the control. At the same time, SNAP caused a decrease in the contraction amplitude, which is in agreement with the known data [20]. Such interaction may be at a level of regulation of both the activities of NO- and H<sub>2</sub>S-synthesizing enzymes and the signaling pathways triggered by these gasses. In vascular tissue, H<sub>2</sub>S reduced the activity of NO synthase [35], while the NO donor, sodium nitroprusside, increased the expression of cystathionin-ylyase and cystathionin- $\beta$ -synthase [36]. In our studies, the effect of NaHS was completely preserved under the conditions of NO synthase inhibition by the nonspecific blocker L-NAME, indicating the absence of NaHS effect on NO synthesis. The interrelationship between H<sub>2</sub>S and NO functions has been shown in the vascular tissues of warm-blooded animals, where sodium nitroprusside increased the vasorelaxation induced by  $H_2S$  [37], while  $H_2S$  reduced the relaxation effect of NO [38].

The negative inotropic effect of NO in frog myocardium is mediated by increase in the level of cGMP, the target of which is cGMP-dependent phosphodiesterase (phosphodiesterase 2). The activation of phosphodiesterase 2 leads to a decrease in the cAMP level, depression of the Ca current, and reduction of the force of contraction [39]. Owing to co-localization of Ca channels and phosphodiesterase 2, which is present in frog mainly in the membrane fraction, a drastic decrease in cAMP concentration occurs only near the Ca channels [40], ensuring cAMP compartmentalization and regulating the phosphorylation of Ca channels separately from other substrates of protein kinase A [39]. It seems that preliminary application of the NO donor results in the activation of phosphodiesterase 2 and decrease in the cAMP level. Under these conditions, the effect of NaHS was reduced, once again demonstrating the potential involvement of phosphodiesterases in the effect of the gas. In addition, one cannot rule out the possibility of chemical interaction between NO and H<sub>2</sub>S with the formation of nitrosothiols, which will have influence on the concentration and effects of both gasses [41].

Thus, our data show the possibility of endogenous synthesis of  $H_2S$  by cystathionin- $\gamma$ -lyase in frog myocardium. Both endogenous and exogenous  $H_2S$  has a negative inotropic effect. The effect of  $H_2S$  is based on the decrease in cAMP level as a result of reduction of its synthesis or increase of its hydrolysis. As a result, the activity of cAMP-dependent protein kinases and phosphorylation of voltage-dependent L-type Ca channels, the Ca entry into cells and the force of myocardial contraction decrease. This effect may play a key role in regulation of the inotropic function of the heart during the activation of  $\beta$ -adrenoceptors involved in the adrenergic regulation of myocardial contractility.

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## REFERENCES

- Beauchamp R.O. Jr., Bus J., Popp J., Boreiko C., Andjelkovich D. 1984. A critical review of the literature on hydrogen sulfide toxicity. *Crit. Rev. Toxicol.* 13, 25–97.
- Sitdikova G.F., Zefirov A.L. 2006. Gaseous mediators in the nerve system. *Ros. Fiziologicheskiy Zh. im. I.M. Sechenova* (Rus.). 97 (7), 872–882.
- 3. Sitdikova G.F., Zefirov A.L. 2010. Hydrogen sulfide: From the Parisian sewer system to a signaling molecule. *Priroda* (Rus.). 9, 29–37.
- Gerasimova E.V., Sitdikova G.F., Zefirov A.L. 2008. Hydrogen sulfide as an endogenous modulator of mediator release at the neuromuscular synapse of the frog. *Neyrokhimia* (Rus.). 25 (1–2), 138–145.
- Elsey D., Fowkes R., Baxter G. 2010. Regulation of cardiovascular cell function by hydrogen sulfide (H<sub>2</sub>S) cell biochemistry and function. *Cell Biochem. Funct.* 28, 95–106.
- 6. Kimura H. 2010. Hydrogen sulfide: From brain to gut. *Antioxid. Redox Signal.* **12**, 1111–1123.
- Sitdikova G., Weiger T., Hermann A. 2010. Hydrogen sulfide increases calcium-activated potassium (BK) channel activity of rat pituitary tumor cells. *Pfluegers Arch. – Eur. J. Physiol.* 459, 389–397.
- Gadalla M., Snyder S. 2010. Hydrogen sulfide as gasotransmitter. J. Neurochem. 113, 14–26.
- 9. Hermann A., Sitdikova G., Weiger T. 2010. Gase als zellulare Signalstoffe. *Biol. Unserer Zeit.* **40**, 185–193.
- Geng B., Yang J., Qi Y., Zhao J., Pang Y., Du J., Tang C. 2004. H<sub>2</sub>S generated by heart in rat and its effects on cardiac function. *Biochem. Biophys. Res. Commun.* **313** (2), 362–368.
- Yong Q., Pan T., Hu L., Bian J. 2008. Negative regulation of β-adrenergic function by hydrogen sulfide in the rat hearts. *J. Mol. Cell Cardiol.* 44 (4), 701–710.
- Bian J., Yong Q., Pan T., Feng Z., Ali M., Zhou S., Moore P. 2006. Role of hydrogen sulfide in the cardioprotection caused by ischemic preconditioning in the rat heart and cardiac myocytes. *J. Pharm. Exp. Ther.* **316** (2), 670–678.
- 13. Dombkowski R., Russell M., Olson K. 2004. Hydrogen sulfide as an endogenous regulator of vascular smooth muscle tone in trout. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **286**, 678–685.
- Olson K. 2005. Vascular actions of hydrogen sulfide in nonmammalian vertebrates. *Antioxid. Redox Signal.* 7, 804–812.

- Abramochkin D.V., Moiseyenko L.S., Kuzmin V.S. 2009. The effect of hydrogen sulfide on electrical activity of rat atrial myocardium. *Byull. Exp. Biol. Meditsiny* (Rus.). 147 (6), 617–621.
- Sun Y., Cao Y., Wang W., Ma S., Yao T., Zhu Y. 2008. Hydrogen sulfide is an inhibitor of L-type calcium channels and mechanical contraction in rat cardiomyocites. *Cardiovasc. Res.* **79** (4), 632–641.
- Xu M., Wu Y., Li Q., Wang F., He R. 2007. Electrophysiological effects of hydrogen sulfide on guinea pig papillary muscles in vitro. *Sheng Li Xue Bao.* 59 (2), 215–220.
- Sitdikova G.F., Khaertdinov N.N., Zefirov A.L. 2011. Investigation of the role of calcium and potassium channels in hydrogen sulfide effects on myocardial contractility in frog. *Byull. Exp. Biol. Meditsiny* (Rus.). 151 (2), 124–128.
- 19. Kimura H. 2000. Hydrogen sulfide induces cyclic AMP and modulates the NMDA receptor. *Biochem. Biophys. Res. Commun.* **267**, 129–133.
- 20. Chesnais J., Fischmeister R., Méry P. 1999. Positive and negative inotropic effects of NO donors in atrial and ventricular fibres of the frog heart. *J. Physiol.* **518**, 449–461.
- 21. Dominy J., Stipanuk M. 2004. New roles for cysteine and transsulfuration enzymes: production of  $H_2S$ , a neuromodulator and smooth muscle relaxant. *Nutr. Rev.* **62** (9), 348–353.
- 22. Teague B., Asiedu S., Moore P. 2002. The smooth muscle relaxant effect of hydrogen sulphide in vitro: evidence for a physiological role to control intestinal contractility. *Br. J. Pharmacol.* **137** (2), 139–145.
- 23. Cheng Y., Ndisang J., Tang G., Cao K., Wang R. 2004. Hydrogen sulfide-induced relaxation of resistance mesenteric artery beds of rats. *Am. J. Physiol. Heart Circ. Physiol.* **287** (5) H2316–2323.
- 24. Zefirov A.L., Sitdikova G.F. 2010. *Ionnye kanaly vozbudimoy kletki (struktura, funktsiya, patologiya)* (Ion Channels of Excitable Cell (Structure, Function, Pathology)). Kazan: Art-kafe.
- 25. Kamp T., Hell J. 2000. Regulation of cardiac L-type calcium channels by protein kinase A and protein kinase C. *Circ. Res.* **87**, 1095–1102.
- Tijskens P., Meissner G., Franzini-Armstrong C. 2003. Location of ryanodine and dihydropyridine receptors in frog myocardium. *Biophys. J.* 84, 1079–1092.
- 27. Odnoshivkina Yu.G., Petrov A.M., Zefirov A.L. 2011. The effect of activation of  $\beta$ 2-adrenoceptors in mouse atriums on the force of contraction, Ca signals, and nitric oxide production. *Acta Naturae* (Rus.). **3**(1), 85– 94.
- Sandberg M., Butt E., Nolte C., Fischer L., Halbrügge M., Beltman J., Jahnsen T., Genieser H., Jastorff B., Walter U. 1991. Characterization of Sp-5,6-dichloro-1-beta-D-ribofuranosylbenzimidazole-3',5'-monophosphorothioate (Sp-5,6-DCl-cBiMPS) as a potent and specific activator of cyclic-AMP-dependent protein

kinase in cell extracts and intact cells. *Biochem. J.* **279** (2), 521–527.

- 29. Sugita S., Baxter D., Byrne J. 1994. cAMP-independent effects of 8-(4-parachlorophenylthio)-cyclic AMP on spike duration and membrane currents in pleural sensory neurons of Aplysia. *J. Neurophysiol.* **72** (3), 1250–1259.
- Connolly B., Willits P., Warrington B., Murray K. 1992. 8-(4-Chlorophenyl)thio-cyclic AMP is a potent inhibitor of the cyclic GMP-specific phosphodiesterase (PDE VA). *Biochem. Pharmacol.* 44 (12), 2303–2306.
- Keef K., Hume J., Zhong J. 2001. Regulation of cardiac and smooth muscle Ca<sup>2+</sup> channels (Ca(V)1.2a,b) by protein kinases. *Am. J. Physiol. Cell Physiol.* 281 (6), C1743–1756.
- Zaccolo M. 2009. cAMP signal transduction in the heart: Understanding spatial control for the development of novel therapeutic strategies. *Br. J. Pharmacol.* 158, 50–60.
- 33. Jurevicius J., Skeberdis V., Fischmeister R. 2003. Role of cyclic nucleotide phosphodiesterase isoforms in cAMP compartmentation following  $\beta$ 2-adrenergic stimulation of ICa,L in frog ventricular myocytes. *J. Physiol.* **551** (1), 239–252.
- Sitdikova G.F., Gerasimova E.V., Khaertdinov N.N., Zefirov A.L. 2009. The role of cyclic nucleotides in the effects of hydrogen sulfide on mediator release in the neuromuscular synapse of frog. *Neyrokhimia* (Rus.). 26 (4), 1–7.
- Kubo S., Doe I., Kurokawa Y., Nishikawa H., Kawabata A. 2007. Direct inhibition of endothelial nitric oxide synthase by hydrogen sulde: Contribution to dual modulation of vascular tension. *Toxicology*. 232, 138– 146.
- 36. Zhao W., Wang R. 2002. H<sub>2</sub>S-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am. J. Physiol. Heart Circ. Physiol.* **283**, 474–480.
- 37. Hosoki R., Matsuki N., Kimura H. 1997. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochem. Biophys. Res. Commun.* 237, 527–531.
- Zhao W., Ndisang J., Wang R. 2003. Modulation of endogenous production of H<sub>2</sub>S in rat tissues. *Can. J. Physiol. Pharmacol.* 81, 848–853.
- Dittrich M., Jurevicius J., Georget M., Rochais F., Fleischmann B., Hescheler J., Fischmeister R. 2001. Local response of L-type Ca<sup>2+</sup> current to nitric oxide in frog ventricular myocytes. *J. Physiol.* 534 (1), 109–121.
- Lugnier C., Gauthier C., Le Bec A., Soustre H. 1992. Cyclic nucleotide phosphodiesterases from frog atrial fibers: Isolation and drug sensitivities. *Am. J. Physiol.* 262, 654–660.
- 41. Whiteman M., Li L., Kostetski I., Chu S., Siau J., Bhatia M., Moore P. 2006. Evidence for the formation of a novel nitrosothiol from the gaseous mediators nitric oxide and hydrogen sulphide. *Biochem. Biophys. Res. Commun.* **343**, 303–310.