## MedChemComm

Cite this: Med. Chem. Commun., 2012, 3, 1449

www.rsc.org/medchemcomm

# 

### Unfolded and macrocyclic ammonium derivatives of diterpenoids steviol and isosteviol having choline moieties. Synthesis and inhibitory activities toward acetylcholine- and butyrylcholinesterases

Mayya G. Korochkina, Alexandra D. Nikitashina, Ravil N. Khaybullin, Konstantin A. Petrov, Irina Yu. Strobykina, Vladimir V. Zobov and Vladimir E. Kataev\*

*Received 25th June 2012, Accepted 9th September 2012* DOI: 10.1039/c2md20165h

A series of unfolded and macrocyclic ammonium derivatives of diterpenoids isosteviol **2** and steviol **17** possessing choline moieties have been synthesized and assayed for inhibitory activities toward AchE and BchE. Compounds **5–8**, **13**, **16**, **20**, and **21** showed moderate activity within the range of IC<sub>50</sub> values  $8.0 \times 10^{-4}$  to  $2.2 \times 10^{-6}$  mol L<sup>-1</sup>. Isosteviol derivative **16** exhibited the best inhibition selectivity against acetylcholinesterase among the compounds tested. It inhibited AchE of human erythrocytes at IC<sub>50</sub> = 80  $\mu$ M, whereas inhibition of BchE occurred at IC<sub>50</sub> > 1000  $\mu$ M.

#### Introduction

Natural products (NPs) are the most consistently successful source of chemicals leading to the development of many drugs for diverse indications, both historically and currently.<sup>1</sup> Every year many new natural products in themselves, semi-synthetic and NP-derived compounds begin to undergo clinical evaluation or registration in the area of diseases, *i.e.* infectious (bacterial, fungal, parasitic and viral), immunological, cardiovascular, neurological, inflammatory and related diseases as well as oncology.<sup>2</sup> Functionalization of naturally occurring compounds with known pharmacophore moieties<sup>3,4</sup> and synthesis of their hybrid compounds<sup>5</sup> are among the most widely used in medicinal chemistry approaches for obtaining novel therapeutic agents.

*Ent*-beyeran diterpenoid isosteviol **2** (ref. 6a) (*ent*-16-oxobeyeran-19-oic acid) is ranked among the most perspective naturally occurring scaffolds for affording novel bioactive compounds. It possesses antihypertension,<sup>6b</sup> hypotension,<sup>6c,d</sup> antihyperglycemic,<sup>6e-g</sup> insulinotropic,<sup>6g</sup> glucanostatic,<sup>6g</sup> antiproliferation,<sup>6h</sup> cardio-<sup>6h,i</sup> and neuroprotective<sup>6j</sup> effects, prevents the growth of cancer cells<sup>6k,d</sup> and exhibits tuberculostatic activity.<sup>6m</sup> About 100 isosteviol derivatives of various structures have been covered in the literature,<sup>7</sup> many of them being bioactive.<sup>8</sup> For example, lactone derivatives of isosteviol demonstrate antiinflammatory activity;<sup>8a</sup> functionalization of isosteviol by introducing a double bond at C15,<sup>8b</sup> by interconversions in ring D<sup>8c</sup> and by coupling of two isosteviol molecules with an amide linker<sup>8d</sup> led to novel *ent*-beyeran-type anticancer agents,<sup>8b-d</sup> functionalization of isosteviol with azine, hydrazide, hydrazone moieties provided a new type of antituberculosis agents.<sup>3a</sup> Recently diterpenoid isosteviol 2 was furnished with a choline moiety Me<sub>3</sub>N<sup>+</sup>-CH<sub>2</sub>CH<sub>2</sub>-O- and the obtained derivatives exhibited high antimicrobial activity.<sup>3b,c</sup> It is known that alkylammonium compounds represent a considerable class of selective reversible inhibitors of cholinesterase.9 We considered that it would be interesting to investigate whether ammonium derivatives of diterpenoid isosteviol having acetylcholine-like structure in which an acyl group is replaced with bulky isostevioyl moiety show anti-cholinesterase activity. In our opinion, the approach to design novel inhibitors of AchE (BchE) on the basis of naturally occurring terpenoids functionalized by choline moiety or even only ammonium group seems to be original for the following reasons. Firstly, the literature provides only a few papers concerning semisynthetic terpenoids having choline moieties. These are the derivatives of abietic,<sup>3d</sup> (+)-dehydroabietic,<sup>3e-g</sup> betulinic,<sup>4b</sup> oleanolic,<sup>4a,b</sup> and ursolic<sup>4b</sup> acids, but in none of the cases anticholinesterase activity is evaluated. Secondly, no information about macrocycles consisting of terpenoid units connected by linkers having quaternary nitrogens is found in the literature. Many macrocycles consisting of ammonium groups connected by polymethylene,10 polyoxamethylene11 (azacoronands) linkers as well as azacyclophanes<sup>12</sup> and azapyrimidinophanes<sup>13</sup> having quaternary nitrogens are known, but in none of the cases anticholinesterase activity was evaluated.

In the present work we describe the synthesis of a series of isosteviol derivatives having one or two choline and *ent*-beyeran moieties in particular unfolded compounds **5–8**, macrocyclic compound **12**, ammonium compound **16** possessing a triethylammonium group coupled with isosteviol skeleton by a tri(oxyethylene) linker, and *ent*-kauranes **20** and **21** which are analogous to *ent*-beyeranes **5** and **6**. The inhibitory activities of some synthesized compounds toward AchE and BchE are also discussed.

A.E. Arbuzov Institute of Organic and Physical Chemistry, Russian Academy of Sciences, Arbuzov str., 8, Kazan, 420088, Russian Federation. E-mail: kataev@iopc.ru; Fax: +7-843-273-2253; Tel: +7-843-231-9160

#### **Results and discussion**

Isosteviol derivatives **5** and **6** having one choline moiety were synthesized as shown in Scheme 1 according to the procedure previously described.<sup>3b</sup> Isosteviol **2** prepared by acid hydrolysis of sweetener Sweta **1** (ref. 14) as previously described<sup>15</sup> was converted to the isosteviol chloride **3** which was further treated with 2-dimethylaminoethanol to give amine **4**. The reaction of amine **4** with alkyl halides afforded ammonium derivatives **5** and **6** in 80% and 70% yields. Then amine **4** was treated with dibromoalkanes according to the procedure previously described<sup>3e</sup> to give ammonium compounds **7** and **8** in 30% and 45% yields. Macrocycle **13** having two *ent*-beyeran (isosteviol) skeletons and two choline moieties was synthesized from isosteviol **2** as shown

in Scheme 1. Initially selective reduction of the isosteviol oxo group with NaBH<sub>4</sub> (ref. 16) gave compound **9** in good yield (92%). Treatment of **9** with sebacoyl chloride in CH<sub>2</sub>Cl<sub>2</sub> in the presence of Py and DMAP furnished **10** in 58% yield.<sup>17</sup> Then diacid **10** was converted with the excess of thionyl chloride into the corresponding acid dichloride **11** in a quantitative yield. The amination of **11** with N,N'-dimethylaminoethanol afforded diamine **12** in 22% yield. Then treatment of **12** with 1,9-dibromononane in CH<sub>3</sub>CN furnished macrocycle **13** in 6% yield.

The macrocyclization hardly proceeded with low conversion of starting amine **12**. When the reaction is completed the reaction mixture was concentrated under vacuum, the obtained crude material was separated by column chromatography into three fractions: macrocycle **13** in 6% yield, starting amine **12** in roughly



Scheme 1 Reagents and conditions: (i) 25%  $H_2SO_4$ ; (ii) SOCl<sub>2</sub>, 40 °C; 2 h; (iii) HOCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>2</sub>, excess, CH<sub>2</sub>Cl<sub>2</sub>, reflux, 21 h; (iv) RX, CH<sub>3</sub>CN, reflux; (v) Br(CH<sub>2</sub>)<sub>n</sub>Br, CH<sub>3</sub>CN, reflux; (vi) NaBH<sub>4</sub>, CH<sub>3</sub>OH, rt; (vii) ClOC(CH<sub>2</sub>)<sub>8</sub>COCl, CH<sub>2</sub>Cl<sub>2</sub>, DMAP, Py; (viii) Br(CH<sub>2</sub>)<sub>9</sub>Br, CH<sub>3</sub>CN, reflux; (ix) Ts(OCH<sub>2</sub>CH<sub>2</sub>)<sub>3</sub>OTs, CH<sub>3</sub>CN, CsCO<sub>3</sub>; (x) LiBr, acetone and (xi) Et<sub>3</sub>N, CH<sub>3</sub>CN.



Scheme 2 Reagents and conditions: (i) NaIO<sub>4</sub>, KOH, H<sub>2</sub>O; (ii) NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O, KOH, H<sub>2</sub>O, RANEY® Ni; (iii) SOCl<sub>2</sub>, 40 °C; 2 h; (iv) HOCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>2</sub>, excess, CH<sub>2</sub>Cl<sub>2</sub>, reflux, 21 h and (iv) RX, CH<sub>3</sub>CN, reflux.

80% yield, and a mixture of unidentified products located near the origin of TLC plate (roughly 10% yield).

It should be noted that even ammonium derivatives of isosteviol **5–8** and **12** are restrictedly dissolved in water owing to their lipophilic hydrocarbon skeletons. To enhance the solubility a polymethylene linker bound ammonium group with isosteviol skeleton was replaced by tri(oxyethylene) one. The first water soluble isosteviol derivative **16** was synthesized as shown in Scheme 1. Alkylation of isosteviol **2** with 1,10-ditosyl-1,4,7,10tetraoxadecane afforded tosylat **14** which was treated with LiBr in acetone to replace the TsO group with bromine. Then the reaction of bromide **15** with Et<sub>3</sub>N in CH<sub>3</sub>CN gave salt **16** in 57% yield.

To evaluate whether the change of the geometry of diterpenoid skeleton from *ent*-beyeran-type (as in isosteviol 2) to *ent*-kauran-type (as in its isomer, diterpenoid steviol 17 (ref. 6a and 18)) modifies inhibitory activity, *ent*-kauranes 20 and 21 having choline moiety were synthesized from 16(S)-dihydrosteviol 18 (ref. 15) as shown in Scheme 2.

The inhibitory activities of compounds **5–8**, **13**, **16**, **20**, and **21** toward human acetyl- (hAchE, EC 3.1.1.7) and butyrylcholinesterase (hBchE, EC 3.1.1.8) were assayed by the spectrophotometric method of Ellman.<sup>19,20</sup> The IC<sub>50</sub> values and selectivity indices (the IC<sub>50</sub> BChE/IC<sub>50</sub> AChE ratios) of compounds synthesized along with the reference selective AchE inhibitor BW284C51 are listed in Table 1.

The analysis of the obtained data showed that all examined compounds exhibited moderate activities within the range of  $IC_{50}$  values  $10^{-4}$  to  $10^{-6}$  mol  $L^{-1}$ , however some interesting regularities can be revealed. Firstly, the worst inhibition activity

**Table 1** Inhibitory potency of tested compounds towardcholinesterases $^{a}$ 

	Compound								
	BW284C51	5	6	7	8	13	16	20	21
AchE (human erythrocytes; Sigma)	0.03 (ref. 21)	150.0	30.0	88	2.2	5.0	80	34	9.0
BchE (human serum; Sigma)	354.0 (ref. 21)	30.0	6.7	65	10	60	>1000	4.2	1.6
SI <sup>b</sup>	11 800	0.2	0.2	0.7	4.5	12	>12.5	0.1	0.2
$^a$ Inhibitory concentrations (IC_{50}) in $\mu M.$ $^b$ Selectivity indices (the IC_{50} BChE/IC_{50} AChE ratio).									

against AchE of human erythrocytes was demonstrated by isosteviol derivative 5 having one choline moiety whereas the best inhibition activity was shown by isosteviol derivatives 8 and 13 having two choline moieties. Unfolded compound 8 was formed from two isosteviol molecules coupled by the linker consisting of two choline moieties connected by a dodecamethylene chain. Macrocycle 13 was formed from two isosteviol molecules coupled by two linkers, namely, (1) the similar linker consisting of two choline moieties connected by a nonamethylene chain. and (2) a diester linker on the basis of sebacic acid. So the coupling of two isosteviol derivatives having one choline moiety results in the increase of inhibition activity against AchE of human erythrocytes. Secondly, isosteviol derivatives 7 and 8 differ just in the length of polymethylene linker between choline moieties. Compound 7 having decamethylene linker exhibited a worse inhibition activity than compound 8 having a dodecamethylene linker. So the lengthening of polymethylene linker between choline moieties considerably increases inhibition activity against AchE of human erythrocytes. Thirdly, the change of the geometry of diterpenoid skeleton affects anticholinesterase activity. Ent-kauranes (steviol derivatives) 20, 21 possess 10 times greater activities against AchE of human erythrocytes and BchE, than their ent-beyeran analogues (isosteviol derivatives) 5, 6. Fourthly, isosteviol derivatives 13 and especially 16 exhibited the highest selectivity toward hAchE among the compounds tested.

#### Conclusions

A series of unfolded and macrocyclic ammonium derivatives of diterpenoids isosteviol **2** and steviol **17** possessing choline moieties have been synthesized and assayed for inhibitory activities toward AchE and BchE. Compounds **5–8**, **13**, **16**, **20** and **21** showed moderate activity within the range of IC<sub>50</sub> values  $8.0 \times 10^{-4}$  to  $2.2 \times 10^{-6}$  mol L<sup>-1</sup>. Isosteviol derivative **16** exhibited the best inhibition selectivity against acetylcholinesterase among the compounds tested. It inhibited AchE of human erythrocytes at IC<sub>50</sub> = 80 µM, whereas inhibition of BchE occurred at IC<sub>50</sub> > 1000 µM.

#### Experimental

#### General

All reagents used were of analytical grade. Solvents were dried if necessary by standard methods. Column chromatography was performed on silica gel (particle size 63–200, mm) using light petroleum–EtOAc as eluent. NMR experiments were carried out with a Bruker AVANCE-600 spectrometer (14.1 T) equipped with a pulsed gradient unit capable of producing magnetic field pulse gradients in the z-direction of 56 G cm<sup>-1</sup>. All spectra were acquired in a 5 mm inverse probehead working at 600.13 MHz in <sup>1</sup>H experiments. Chemical shifts are reported on the  $\delta$  (ppm) scale and are relative to the residual <sup>1</sup>H signal of CDCl<sub>3</sub>.

**Dichloride 11.** Diacid **10** (0.9 g, 1.1 mmol) was dissolved in a large excess SOCl<sub>2</sub> and heated for 2 h at 40 °C. The reaction mixture was concentrated under vacuum to afford **11** as viscous oil. Yield: 0.94 g (100%). IR spectrum ( $\nu$ /cm<sup>-1</sup>): 1732, 1795 (C=O). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm), *J* (Hz): 0.79–2.00 (m, 50H, *ent*-beyeran skeleton and –(CH<sub>2</sub>)<sub>6</sub>–); 0.82 (s, 6H, C<sup>20</sup>'H<sub>3</sub>, C<sup>20</sup>H<sub>3</sub>); 0.91 (s, 6H, C<sup>17'</sup>H<sub>3</sub>, C<sup>17</sup>H<sub>3</sub>); 1.32 (s, 6H, C<sup>18'</sup>H<sub>3</sub>, C<sup>18</sup>H<sub>3</sub>); 2.32 (m, 6H, C<sup>3'</sup>H<sub>eq</sub>, C<sup>3</sup>H<sub>eq</sub> and C<sup>16'</sup>, C<sup>16</sup>–OC(O)CH<sub>2</sub>CH<sub>2</sub>); 4.75 (dd, 2H, *J* = 10.5, 4.3, C<sup>16'</sup>H, C<sup>16</sup>H).

**Diamine 12.** The solution of dichloride **11** in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was added dropwise to a stirred solution of *N*,*N*-dimethylethanolamine (5 mL) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) under argon. The mixture was refluxed for 21 h under argon. After that the reaction mixture was washed with saturated NaHCO<sub>3</sub> aqueous solution, the organic layer was dried with CaCl<sub>2</sub> and concentrated under vacuum. The residue was purified by column chromatography on silica (chloroform–methanol = 6 : 1, v/v) to give diamine **12** (0.26 g, 22%) as viscous oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.69–2.11 (m, 50H, *ent*-beyeran skeleton and –(CH<sub>2</sub>)<sub>6</sub>–), 0.69 (s, 6H, C<sup>20</sup>H<sub>3</sub>, C<sup>20</sup>H<sub>3</sub>), 0.88 (s, 6H, C<sup>17′</sup>H<sub>3</sub>, C<sup>17</sup>H<sub>3</sub>), 1.14 (s, 6H, C<sup>18′</sup>H<sub>3</sub>, C<sup>18</sup>H<sub>3</sub>), 2.14 (d, 2H, C<sup>3′</sup>H<sub>eq</sub>, C<sup>3</sup>H<sub>eq</sub>, *J* = 13.3 Hz), 2.25 (s, 12H, –N(CH<sub>3</sub>)<sub>2</sub>), 2.28 (t, 2H, C<sup>16′</sup>,C<sup>16</sup>–OC(O)CH<sub>2</sub>CH<sub>2</sub>, *J* = 7.5 Hz), 2.53 (m, 4H, –C(O)OCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>2</sub>), 4.10 (m, 4H, C(O) OCH<sub>2</sub>–), 4.70 (dd, 2H, C<sup>16′</sup>H, C<sup>16′</sup>H, *J* = 10.3, 4.6 Hz). MALDI-TOF MS, *m/z*: 950 [M + H]<sup>+</sup>; *m/z* (calc.): 948.72 (C<sub>58</sub>H<sub>96</sub>N<sub>2</sub>O<sub>8</sub>).

Macrocycle 13. The solution of 1,9-dibromononane (0,10 mL, 0.49 mmol) in CH<sub>3</sub>CN (15 mL) was added to a stirred solution of 12 (0.26 g, 0.27 mmol) in CH<sub>3</sub>CN (100 mL) and the reaction mixture was refluxed for 50 h under argon. After that the reaction mixture was concentrated under vacuum, and the residue was purified by column chromatography on silica (chloroformmethanol = 2: 1, v/v) to give macrocycle 13 (0.02 g, 6.0%), white powder, mp 194–198 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.66-2.10 (m, 64H, ent-beyeran skeleton, -(CH<sub>2</sub>)<sub>6</sub>- and -(CH<sub>2</sub>)<sub>7</sub>-), 0.66 (s, 6H,  $C^{20'}H_3$ ,  $C^{20}H_3$ ), 0.90 (s, 6H,  $C^{17'}H_3$ ,  $C^{17}H_3$ ), 1.18 (s, 6H,  $C^{18'}H_3$ ,  $C^{18}H_3$ ), 2.12 (d, 2H,  $C^{3'}H_{eq}$ ,  $C^{3}H_{eq}$ , J = 13.3 Hz), 2.32 (m, 2H,  $C^{16'}$ ,  $C^{16}$ –OC(O)CH<sub>2</sub>CH<sub>2</sub>), 3.41 (s, 12H, -N(CH<sub>3</sub>)<sub>2</sub>), 3.89 (m, 4H, -NCH<sub>2</sub>(CH<sub>2</sub>)<sub>7</sub>CH<sub>2</sub>N-), 4.07 (m, 4H, -C(O) OCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>2</sub>-), 4.45 and 4.63 (m, 4H, C(O)OCH<sub>2</sub>-), 4.68  $(dd, 2H, C^{16'}H, C^{16}H, J = 10.3, 4.6 Hz)$ . MALDI-TOF MS, *m/z*:  $1060 [M - CH_3]^+$ ,  $1154 [M - Br]^+$ .  $1228 [M-DHB + H]^+$ . Elemental anal. C 65.43; H 8.97; Br 13.09; N 2.35, calcd for C<sub>67</sub>H<sub>114</sub>Br<sub>2</sub>N<sub>2</sub>O<sub>8</sub>, C 65.14; H 9.30; Br 12.94; N 2.27%.

**Tosylate 14.**  $\alpha$ , $\omega$ -Ditosyl triethylene glycol (10.8 g, 23.5 mmol) was added to a stirred solution of isosteviol **2** (5.0 g, 15.7 mmol) in CH<sub>3</sub>CN (250 mL). Then the reaction mixture was heated up to 70 °C and K<sub>2</sub>CO<sub>3</sub> (3.3 g, 23.8 mmol) was added. After heating for 6 h at 70 °C the precipitate was filtered and the mixture was

concentrated under vacuum. The crude product was purified by column chromatography on silica (petroleum ether 40–70%– ethyl acetate = 2 : 1, v/v) to give tosylate **14** (5.0 g, 53%) as viscous oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.62–2.14 (m, 18H, *ent*-beyeran skeleton), 0.70 (s, 3H, C<sup>20</sup>H<sub>3</sub>), 0.96 (s, 3H, C<sup>17</sup>H<sub>3</sub>), 1.18 (s, 3H, C<sup>18</sup>H<sub>3</sub>), 2.17 (d, 1H, C<sup>3</sup>H<sub>eq</sub>, J = 13.2 Hz), 2.62 (dd, 1H, C<sup>15</sup>H<sub>a</sub>, J = 3.4, 19.1 Hz), 3.56 (brs, 4H, OCH<sub>2</sub>-CH<sub>2</sub>O), 3.64 (t, 2H, C(O)OCH<sub>2</sub>CH<sub>2</sub>O–, J = 4.7 Hz), 3.68 (t, 2H, –OCH<sub>2</sub>CH<sub>2</sub>OTs, J = 4.7 Hz), 4.15 (m, 4H, C(O)OCH<sub>2</sub>– and –CH<sub>2</sub>OTs). MALDI-TOF MS, *m*/*z*: 627 [M + Na]<sup>+</sup>, 643 [M + K]<sup>+</sup>. Elemental anal. C 65.56; H 8.24; S 5.16, calcd for C<sub>33</sub>H<sub>48</sub>O<sub>8</sub>S, C 65.54; H 8.00; S 5.30%.

Bromide 15. Lithium bromide (0.79 g, 9.1 mmol) was added to a stirred solution of tosylate 14 (5.0 g, 8.3 mmol) in acetone (25 mL). The reaction mixture was refluxed for 16 h. After that the precipitate was filtered and the mixture was concentrated under vacuum. The crude product was purified by column chromatography on silica (petroleum ether-ethyl acetate = 2:1, v/v) to give bromide 15 (3.7 g, 88%) as viscous oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz), δ (ppm): 0.71-2.15 (m, 18H, ent-beyeran skeleton), 0.71 (s, 3H, C<sup>20</sup>H<sub>3</sub>), 0.97 (s, 3H, C<sup>17</sup>H<sub>3</sub>), 1.19 (s, 3H,  $C^{18}H_3$ ), 2.19 (d, 1H,  $C^{3}H_{eq}$ , J = 13.5 Hz), 2.62 (dd, 1H,  $C^{15}H_a$ , J = 3.7, 18.6 Hz), 3.45 (t, 2H,  $-CH_2$ Br, J = 6.3 Hz), 3.64 (m, 4H, OCH<sub>2</sub>CH<sub>2</sub>O), 3.69 (m, 2H, -OCH<sub>2</sub>CH<sub>2</sub>Br), 3.80 (t, 2H, C(O) OCH<sub>2</sub>CH<sub>2</sub>O-, J = 6.2 Hz), 4.18 (m, 2H, C(O)OCH<sub>2</sub>-). MALDI-TOF MS, m/z: 537 [M + Na]<sup>+</sup>, 552 [M + K]<sup>+</sup>. Elemental anal. C 60.27; H 8.09; Br 15.45, calcd for C<sub>26</sub>H<sub>41</sub>BrO<sub>5</sub>, C 60.81; H 8.05; Br 15.56%.

**Compound 16.** Triethylamine (9.0 mL, 66.2 mmol) was added to a stirred solution of bromide **15** (3.4 g, 6.6 mmol) in CH<sub>3</sub>CN (80 mL). The reaction mixture was refluxed for 17 h, concentrated under vacuum, and the crude product was purified by column chromatography on silica (chloroform/methanol) to give compound **16** (2.3 g, 57%) as a white powder, mp 90–93 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.67–2.02 (m, 18H, *ent*beyeran skeleton), 0.67 (s, 3H, C<sup>20</sup>H<sub>3</sub>), 0.94 (s, 3H, C<sup>17</sup>H<sub>3</sub>), 1.16 (s, 3H, C<sup>18</sup>H<sub>3</sub>), 1.37 (t, 9H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>3</sub>, J = 7.2), 2.14 (d, 1H, C<sup>3</sup>H<sub>eq</sub>, J = 13.2 Hz), 2.56 (dd, 1H, C<sup>15</sup>H<sub>a</sub>, J = 3.6, 18.4 Hz), 3.54 (m, 8H, –CH<sub>2</sub>N(CH<sub>2</sub>CH<sub>3</sub>)<sub>3</sub>), 3.64 (m, 4H, OCH<sub>2</sub>CH<sub>2</sub>O), 3.75 (m, 2H, OCH<sub>2</sub>), 3.95 (m, 2H, C(O)OCH<sub>2</sub>CH<sub>2</sub>O), 4.13 (m, 2H, C(O) OCH<sub>2</sub>–). MALDI-TOF MS, *m*/*z*: 534 [M – Br]<sup>+</sup>. Elemental anal. C 62.43; H 9.30; N 2.37; Br 13.19, calcd for C<sub>32</sub>H<sub>56</sub>Br NO<sub>5</sub>, C 62.53; H 9.18; Br 13.01; N 2.28%.

#### General procedure for synthesis of compounds 20 and 21

Amine 19. 13-Dihydrosteviol 18 (0.5 g, 1.6 mmol) was dissolved in a large excess SOCl<sub>2</sub> and heated for 3 h at 40 °C. The reaction mixture was concentrated under vacuum affording appropriate dichloride of hydroxyacid 18 as viscous oil. A solution of crude product (0.4 g, 1 mmol) in CCl<sub>4</sub> (30 mL) was added dropwise to a stirred solution of N,N-dimethylethanoloamine (2 mL, 20 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) under argon and the mixture was refluxed for 24 h under argon. After that the reaction mixture was washed with saturated NaHCO<sub>3</sub> aqueous solution, the organic layer was dried with CaCl<sub>2</sub> and concentrated under vacuum. The residue was purified by column chromatography on silica (chloroform–methanol = 9 : 1, v/v) to give amine **19** (0.25 g, 54%) as viscous oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.72–2.24 (m, 20H, *ent*-kaurane skeleton), 0.83 (s, 3H, C<sup>20</sup>H<sub>3</sub>), 1.03 (s, 3H, C<sup>17</sup>H<sub>3</sub>), 1.13 (s, 3H, C<sup>18</sup>H<sub>3</sub>), 2.25 (s, 6H, -N(CH<sub>3</sub>)<sub>2</sub>), 2.33 (m, 1H), 2.54 (m, 2H, -C(O) OCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>2</sub>), 4.10 (m, 2H, C(O)OCH<sub>2</sub>–). MALDI-TOF MS, *m*/*z*: 410 [M]<sup>+</sup>.

Alkyl halide (0.6 mmol) was added to a stirred solution of amine **19** (0.6 mmol) in CH<sub>3</sub>CN (5 mL). After heating for 10 h at 80 °C, the mixture was concentrated under vacuum and the crude product was recrystallized from hexane.

**Compound 20.** Yield 55%; mp 226–228 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.78–2.32 (m, 21H, *ent*-kaurane skeleton), 0.82 (s, 3H, C<sup>20</sup>H<sub>3</sub>), 1.06 (s, 3H, C<sup>17</sup>H<sub>3</sub>), 1.21 (s, 3H, C<sup>18</sup>H<sub>3</sub>), 3.57 (s, 9H, –N(CH<sub>3</sub>)<sub>3</sub>), 4.11 (m, 2H, –C(O)OCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>3</sub>), 4.55 (m, 2H, C(O)OCH<sub>2</sub>–). MALDI-TOF MS, *m/z*: 424 [M – I]<sup>+</sup>. Elemental anal. C 54.12; H 7.40; Cl 7.19; I 22.48; N 2.79; calcd for C<sub>25</sub>H<sub>43</sub>ClINO<sub>2</sub>; C 54.40; H 7.85; Cl 6.72; I 22.99; N 2.54%.

**Compound 21.** Yield 34%; mp 190–192 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz),  $\delta$  (ppm): 0.75–2.15 (m, 21H, *ent*-kaurane skeleton), 0.75 (s, 3H, C<sup>20</sup>H<sub>3</sub>), 1.06 (s, 3H, C<sup>17</sup>H<sub>3</sub>), 1.16 (s, 3H, C<sup>18</sup>H<sub>3</sub>), 3.40 (s, 6H, -N(CH<sub>3</sub>)<sub>2</sub>), 4.06 (m, 2H, -C(O)OCH<sub>2</sub>CH<sub>2</sub>N(CH<sub>3</sub>)<sub>3</sub>), 4.57 (m, 2H, C(O)OCH<sub>2</sub>–), 5.20 (s, 2H, -CH<sub>2</sub>Ph), 7.47–7.70 (m, 5H, Ph). MALDI-TOF MS, *m*/*z*: 500 [M – Br]<sup>+</sup>. Elemental anal. C 63.99; H 8.83; Br 14.05; Cl 7.07; N 2.06; calcd for C<sub>31</sub>H<sub>47</sub>BrClNO<sub>2</sub>, C 64.08; H 8.15; Br 13.75; Cl 6.1; N 2.41%.

#### Acknowledgements

We gratefully thank Dr V.M. Babaev (A.E.Arbuzov Institute of Organic and Physical Chemistry, Kazan Sci. Center of RAS, Russia) for MALDI-TOF experiments. This work was supported by funding from Russian Foundation for Basic Research (grant 10-03-00499-a), Presidium of RAS (program no. 8) and Branch of Chemistry and Material Science of RAS (program no. 6).

#### Notes and references

- (a) B. B. Mishra and V. K. Tiwari, *Eur. J. Med. Chem.*, 2011, **46**, 4769;
   (b) J. A. Beutler, *Current Protocols in Pharmacology*, Issue SUPPL., 2009, vol. 46, pp. 9111–9121;
   (c) S. Coseri, *Mini-Rev. Med. Chem.*, 2009, **9**, 560;
   (d) K.-H. Lee, *J. Nat. Prod.*, 2010, **73**, 500.
- 2 (a) D. J. Newman, G. M. Cragg and K. M. Snader, J. Nat. Prod., 2003, **66**, 1022; (b) D. J. Newman and G. M. Cragg, J. Nat. Prod., 2007, **70**, 461; (c) D. J. Newman, J. Med. Chem., 2008, **51**, 2589.
- 3 (a) V. E. Kataev, I. Yu. Strobykina, O. V. Andreeva, B. F. Garifullin, R. R. Sharipova, V. F. Mironov and R. V. Chestnova, Russ. J. Bioorg. Chem., 2011, 37, 483; (b) M. G. Korochkina, R. R. Sharipova, I. Yu. Strobykina, A. V. Lantsova, A. D. Voloshina, N. V. Kulik, V. V. Zobov, V. E. Kataev and V. F. Mironov, Pharm. Chem. J., 2011, 44, 597; (c) M. G. Korochkina, V. M. Babaev, I. Yu. Strobykina, A. D. Voloshina, N. V. Kulik and V. E. Kataev, Chem. Nat. Compd., 2012, 47, 914; (d) N. V. Kryuchkova, Yu. N. Orlov, D. A. Golovacheva and S. V. Levanova, Russ. J. Appl. Chem., 2011, 84, 284; (e) W. Jia, X. Rao, Z. Song and S. Shang, J. Surfactants Deterg., 2009, 12, 261; (f) B. A. Radbil', S. R. Kushnir, A. B. Radbil', E. N. Shmidt, V. F. Smirnov and N. B. Mel'nikova, Russ. J. Appl. Chem., 2008, 81, 874; (g) G.-Y. Cui, B.-R. Mo, W.-N. Chen and K.-R. Huang, Riyong Huaxue Gongye, 2007, 37, 202
- 4 (a) M. I. Hamed and Z. El-Gholmy, *Arzneim. Forsch.*, 1972, **22**, 2133; (b) D. Biedermann, B. Eigenrova, M. Hajduch and J. Sarek, *Synthesis*, 2010, 3839.

- 5 (a) K. Gademann, Chimia, 2006, 60, 841; (b) L. F. Tietze, H. P. Bell and S. Chandrasekhar, Angew. Chem., Int. Ed., 2003, 42, 3996; (c) D. B. Salunke, B. G. Hazra and V. S. Pore, Curr. Med. Chem., 2006, 13, 813; (d) L. Pouységu, D. Deffieux, G. Malik, A. Natangelo and S. Quideau, Nat. Prod. Rep., 2011, 28, 853; (e) O. V. Andreeva, R. R. Sharipova, I. Yu. Strobykina, O. A. Lodochnikova, A. B. Dobrynin, V. M. Babaev, R. V. Chestnova, V. F. Mironov and V. E. Kataev, Chem. Nat. Compd., 2011, 46, 902.
- 6 (a) E. Mosettig and W. R. Nes, J. Org. Chem., 1955, 20, 884; (b) J. Liu, P. Kao, M. Hsieh and Y. Chen, Acta. Cardiol. Sin., 2001, 17, 133; (c) K. L. Wong, P. Chan, H. Y. Yang, F. L. Hsu, I. M. Liu, Y. W. Cheng and J. T. Cheng, *Life Sci.*, 2004, **74**, 2379; (d) K. L. Wong, H. Y. Yang, P. Chan, T. H. Cheng, J. C. Liu, F. L. Hsu, I. M. Liu, Y. W. Cheng and J. T. Cheng, Planta Med., 2004, 70, 108; (e) J. Ma, Z. Ma, J. Wang, R. W. Milne, D. Xu and A. K. Davey, *Diabetes, Obes. Metab.*, 2007, **9**, 597; (*f*) D. Xu, M. Xu, L. Lin, S. Rao, J. Wang and A. K. Davey, *Life Sci.*, 2012, **90**, 30; (*g*) P. B. Jeppesen, S. Gregersen, K. K. Alstrup and K. Hermansen, Phytomedicine, 2002, 9, 9; (h) K. L. Wong, J. W. Lin, J. C. Liu, H. Y. Yang, P. F. Kao, C. H. Chen, S. H. Loh, W. T. Chiu, T. H. Cheng, J. G. Lin and H. Hong, J. Pharmacol., 2006, 76, 163; (*i*) D. Xu, Y. Li, J. Wang, A. K. Davey, S. Zhang and A. M. Evans, *Life Sci.*, 2007, **80**, 269; (*j*) D. Xu, W. Du, L. Zhao, A. K. Davey and J. Wang, Planta Med., 2008, 74, 816; (k) Y. Mizushina, T. Akihisa, M. Ukiya, Y. Hamasaki, C. Murakami-Nakai, I. Kuriyama, I. Kuriyama, T. Takeuchi, F. Sugawara and H. Yoshida, *Life Sci.*, 2005, **77**, 2127; (*l*) M. Takasaki, T. Konoshima, M. Kozuka, H. Tokuda, J. Takayasu, H. Nishino, M. Miyakoshi, K. Mizutani and K.-H. Lee, Bioorg. Med. Chem., 2009, 17, 600; (m) V. E. Kataev, O. I. Militsina, I. Yu. Strobykina, G. I. Kovylyaeva, R. Z. Musin, O. V. Fedorova, G. L. Rusinov, M. N. Zueva, G. G. Mordovskoi and A. G. Tolstikov, Pharm. Chem. J., 2006, 40, 463.
- 7 For a general review on isosteviol chemistry see (a) V. E. Kataev, R. N. Khaybullin, R. R. Sharipova and I. Yu. Strobykina, *Review Journal of Chemistry*, 2011, 1, 93; (b) N. Moons, W. De Borggraeve and W. Dehaen, *Curr. Org. Chem.*, 2011, 15, 2731.
- 8 (a) B.-H. Chou, L.-M. Yang, S.-F. Chang, F.-L. Hsu, C.-H. Lo,
  W.-K. Lin, L.-H. Wang, P.-C. Liu and S.-J. Lin, *Phytochemistry*,
  2009, **70**, 759; (b) J. Li, D. Zhang and X. Wu, *Bioorg. Med. Chem.*Lett., 2011, **21**, 130; (c) Y. Wu, G.-F. Dai, J.-H. Yang,
  Y.-X. Zhang, Y. Zhu and J.-C. Tao, *Bioorg. Med. Chem. Lett.*,
  2009, **19**, 1818; (d) L.-H. Lin, L.-W. Lee, S.-Y. Sheu and P.-Y. Lin,
  Chem. Pharm. Bull., 2004, **52**, 1117.
- 9 K. B. Augustinsson, The Evolution of Esterases in Vertebrates, in Homologous Enzymes and Biochemical Evolution, ed. N. Van Thoai and J. Roche, Gordon & Breach Inc., New York, 1968, pp. 299–311.
- 10 (a) R. Danieli and A. Ricci, J. Chem. Soc., Perkin Trans. 2, 1976, 290;
  (b) F. P. Schmidtchen, Angew. Chem., Int. Ed. Engl., 1977, 16, 720; (c)
  S. I. Lall, D. Mancheno, S. Castro, V. Behaj, J. L. I. Cohen and
  R. Engel, Chem. Commun., 2000, 2413; (d) S. Souirti and
  M. Baboulene, Synth. Commun., 2001, 31, 9.
- (a) J. Jurczak, R. Ostaszewski and P. Satanski, J. Chem. Soc., Chem. Commun., 1989, 184; (b) J. Jurczak, R. Ostaszewski, P. Satanski and T. Stankiewicz, Tetrahedron, 1993, 49, 1471; (c) M. Mori, H. Tsue, K. Tanaka and S. Tanaka, Bull. Chem. Soc. Jpn., 2003, 76, 121.
- (a) Y. Murakami, A. Nakano, R. Miyata and Y. Matsuda, J. Chem. Soc., Perkin Trans. 1, 1979, 1669; (b) G. R. Newkome, V. K. Majestic and F. R. Fronczek, Tetrahedron Lett., 1981, 22, 3039; (c) S. Shinkai, A. Yoshioka, H. Nakayama and O. Manabe, J. Chem. Soc., Perkin Trans. 2, 1990, 1905; (d) H. Takemura, Y. Kozima and T. Inazu, Tetrahedron Lett., 1999, 40, 6431.
- 13 V. E. Semenov, A. D. Voloshina, E. M. Toroptzova, N. V. Kulik, V. V. Zobov, R. K. Giniyatullin, A. S. Mikhailov, A. E. Nikolaev, V. D. Akamsin and V. S. Reznik, *Eur. J. Med. Chem.*, 2006, **41**, 1093.
- 14 Enzymatically treated glycosides of the plant *S. rebaudiana* manufactured by Stevian Biotechnology Corporation (Kuala Lumpur, Malaysia).
- 15 R. N. Khaibullin, I. Yu. Strobykina, V. E. Kataev, O. A. Lodochnikova, A. T. Gubaidullin and R. Z. Musin, *Russ. J. Gen. Chem.*, 2009, **79**, 967.

- 16 V. A. Al'fonsov, G. A. Bakaleinik, A. T. Gubaidullin, V. E. Kataev, G. I. Kovylyaeva, A. I. Konovalov, I. A. Litvinov, I. Yu. Strobykina, S. I. Strobykin, O. V. Andreeva and M. G. Korochkina, *Russ. J. Gen. Chem.*, 2000, **70**, 953.
- 17 Previously diacid 10 was obtained by reflux of hydroxyacid 9 with sebacinoyl dichloride in CCl<sub>4</sub> with CoCl<sub>2</sub> for 15 h in moderate yield<sup>6m</sup> (35%). In order to improve the yield of 10, the reaction condition was optimized in this study as follows. A solution of (0.05 g, 0.4 mmol) DMAP and 0.08 mL (1 mmol) pyridine was added to solution of 1.15 g (3.6 mmol) hydroxyacid 9 in 500 mL CH<sub>2</sub>Cl<sub>2</sub>. Then solution of (0.43 g, 1.8 mmol) sebacinoyl dichloride in 10 mL CH<sub>2</sub>Cl<sub>2</sub> was added dropwise at 0 °C. The mixture was stirred at room temperature for 72 h, washed with acidulous water and NaCl aqueous solution. The extract was dried with MgSO4 and concentrated under vacuum. The crude product was purified by column chromatography on silica (petroleum ether-ethyl acetate = 5 : 1, v/v) to give 0.84 g (58%) of diacid 10 as white powder; mp 108-110 °C (ethanol). IR spectrum (v/cm<sup>-1</sup>): 1181, 1255, 1733 (O-C-O), 1694 (COOH). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz), δ (ppm): 0.76–2.11 (m, 52H, ent-beyeran skeleton and -(CH<sub>2</sub>)<sub>6</sub>-), 0.76 (s, 6H, C<sup>20</sup>H<sub>3</sub>,  $C^{20}$ H3), 0.91 (s, 6H,  $C^{17}$ H<sub>3</sub>,  $C^{17}$ H<sub>3</sub>), 1.20 (s, 6H,  $C^{18}$ H<sub>3</sub>,  $C^{18}$ H<sub>3</sub>), 2.13 (d, 2H, J 13.1, C3Heq, C<sup>3</sup>Heq), 2.33 (m, 4H, C<sup>16</sup>, C<sup>16</sup>–OC(O) CH<sub>2</sub>CH<sub>2</sub>); 4.62 (dd, 2H, J 10.2, 4.3, C<sup>16</sup>H, C<sup>16</sup>H). MALDI-TOF MS, m/z (exp): 830 [M + Na]<sup>+</sup>; m/z (calc.): 806.57 [M]<sup>+</sup> (C<sub>50</sub>H<sub>78</sub>O<sub>8</sub>).
- 18 M. Bridel and R. Lavieille, J. Pharm. Chim., 1931, 14, 99.

- 19 G. L. Ellman, K. D. Courtney, V. Andres Jr and R. M. Feather-Stone, *Biochem. Pharmacol.*, 1961, 7, 88.
- 20 The inhibition assays of AChE from human erythrocytes (Sigma) and BChE from human serum (Sigma) were run in phosphate buffer 0.1 M, at pH 8.0. Acetyl- and butyrylthiocholine iodides were used as substrates and 5,5'-dithiobis(2-nitrobenzoic acid) (DTNB) was used as the chromophoric reagent. Inhibition assays were carried out on a Perkin Elmer  $\lambda 25$ -visiblespectrophotometer. AChE inhibitory activity was determined in the reaction mixture containing 400  $\mu$ L of the solution of AChE (0.415 U mL<sup>-1</sup> in 0.1 M phosphate buffer, pH 8.0), 200 µL of the 3.3 mM solution of DTNB in 0.1 M phosphate buffer (pH 7.0) containing 6 mM NaHCO<sub>3</sub>, 200 µL of inhibitor solution and 1000 µL of phosphate buffer, pH 8.0. After incubation for 20 min at 36 °C, acetylthiocholine iodide (200 µL of 0.05 mM water solution) was added as the substrate and AChE-catalyzed hydrolysis was studied by measuring the increase of absorbance at 412 nm for 1.0 min at 36 °C. Then inhibitor concentration for 50% inhibition of the AChE activity (IC<sub>50</sub>) was calculated by non-linear regression of the response-concentration (log) curve. BChE inhibitory activity was assessed similarly using butyrylthiocholine iodide (0.05 mM) as the substrate.
- 21 K. Musilek, M. Komloova, O. Holas, M. Hrabinova, M. Pohanka, V. Dohnal, F. Nachon, M. Dolezal and K. Kuca, *Eur. J. Med. Chem.*, 2011, 46, 811.